QUALITATIVE AND QUANTITATIVE ANALYSIS OF ORGANOSULPHUR COMPOUNDS IN SELECTED FUEL SAMPLES

by

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Organosulphur compounds, crude oil, diesel, gasoline, kerosene, gas chromatography, timeof-flight mass spectrometry, magnetic solid phase extraction, dichloromethane, nano materials. Above all, I would like to first thank God.

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- S. S. Mgiba, V. Mhuka, N. C. Hintsho-Mbita & Nomvano Mketo, Selective magnetic solid phase extraction based on Au-Fe₃O₄ adsorbent for chromatographic determination of organsulphur compounds in selected fuel oil samples, in preparation.

ABSTRACT

Crude oil is a complex mixture of hydrocarbons, non-hydrocarbons and heteroatomcontaining compounds. This fossil fuel undergoes refining processes to produce different fuel oils, which contributes one third to one-half of the world's energy supply. The presence of organosulphur compounds (OSCs) in fuel oils is undesirable because these compounds emit sulphur oxide (SO_x) gases to the atmosphere, corrode vehicle engines and deactivate catalyst during the refinery processes of crude oil. On the other hand, the identification and quantification of OSCs in fuel oils has been a challenging task. This is due to their volatile nature, the substituents structure of some OSCs such as polycyclic aromatic sulphur heterocycles (PASH) and the presence of other polycyclic hydrocarbons in high concentration, which causes interferences during the chromatographic analysis. Therefore, there has been a progress on development of various analytical methods for both qualitative and quantitative determination of OSCs in fuels.

Therefore, this study reports a rapid and efficient method for the qualitative analysis of OSCs in fuel oil samples. A simple sample dilution with dichloromethane was conducted for qualitative analysis of the OSCs in crude oil, kerosene, gasoline and diesel samples using gas chromatographytime of flight-mass spectrometry (GC-T-TOFMS). Diesel sample showed the largest number of OSCs (26) namely: Thiophene, 3-methyl; 2-Thiophenecarboxylic acid, 4-nitrophenyl ester; Acetic acid, mercapto-; Thiazol-2-amine, N-(4-dimethylaminobenzyl)-; benzo[b]thiophene; Thieno [3,4-d]-1,3-dioxole, tetrahydro-2,2-dimethyl-, 5,5-dioxide; dibenzothiophene Sulfurous acid, octyl 2-propyl ester; Benz(1,4) oxathiino (2,3-c)pyridine; 3-hydroxy-2thiabutane; 2-Thiopheneacetic acid, isopropyl ester; 3H-[1,3,4]Oxadiazole-2-thione, 5-(4-fluorophenyl)-3-(piperidin-1-yl)methyl-; 2-Butanone, 3,3-dimethyl-1-thiocyanato-; 3-Acetyl-2,5-dichlorothiophene; 2(3H)-Thiazolethione, 4-methyl-; 1,3-Diphenyl-4H-1,2,4-triazoline-5-thione; Benzothiazole, 2-(m-tolyl)-; 1,3-Diphenyl-4H-1,2,4-triazoline-5-thione; Ethylamine, N,N-dinonyl-2-phenylthio; 1,3-Diphenyl-4H-1,2,4-triazoline-5-1,3-Diphenyl-4H-1,2,4-triazoline-5-thione; 2thione: 10H-Phenothiazine, (trifluoromethyl)-; 1,3-Diphenyl-4H-1,2,4-triazoline-5-thione; 1-(4-Methyldibenzothiophen-2-yl)-piperidine; 2,2-Dimethyl-propyl 2,2-dimethyl-propanethiosulfinate; Difluoro(methylamino)phosphine sulfide followed by crude oil (10), kerosene (11) and gasoline with 8 OSCs, namely: Thiophene, 2-(1,1-dimethylethyl); 3-Picoline, 2-(tert-butylthiol); 2-Thiophenecarboxylic acid, 4-nitrophenyl ester; methanesulfonamide, N-[2-(octylamino)ethyl]; Tetramethyl diphosphane-oxidesulfide; Sulfurous acid, 2-ethylhexyl hexyl ester; 2-Thiophenecarboxylic acid, 5-tertbutyl and Thiophene, 2-nitro.

The study also describes the development of environmentally friendly, effective, and selective pre-concentration method of OSCs in fuels, followed by GC-TOF-MS for both qualitative and semi-quantitative analysis. The magnetic solid phase extraction (m-SPE) based on Au-Fe₃O₄ adsorbent was selected as the preconcentration method for selected OSCs (thiophene; 3-methylthiophene; benzothiophene and dibenzothiophene). The Au-Fe₃O₄ NPs were synthesized using a co-precipitation method and characterized using XRD which produced diffraction peaks for gold coated magnetite with miller indices of Au (111), (200),(220), (311) and (222). UV-Vis was also used and a surface plasmon resonance peak at around 520 - 523 was produced. TEM and SEM confirmed the spherical shape and the size of 100 nm of the nanoparticles. Lastly, FTIR confirmed the particles with vibrational peaks at 500 cm⁻¹ for Fe-O bond, 1500 cm⁻¹ and 1750 cm⁻¹ for C=O and –OH groups. The most influential parameters (mass of the adsorbent, extraction time, eluent volume and pH of the sample) affecting m-SPE were investigated by using multivariate mathematical tools. The best optimum conditions were found to be 150 mg mass of adsorbent, 100 µL eluent volume of acetonitrile, 50 minutes extraction time and 6,5 pH of the sample. These optimum conditions showed relatively low limits of detection $(0,02-0,199 \mu g/L)$ and limit of quantification $(0,126-0,602 \mu g/L)$ for all four of the investigated compounds. Furthermore, precision of 0.2 - 0.8% was obtained with good linearity of 0.9816-0,9961. The extraction efficiency of the proposed m-SPE method ranged from 76 to 95% for the spiked samples (4- 12 µg/L spike concentration level). The optimized m-SPE method was then applied toreal fuel oil samples and the concentration levels of non-target OSCs in crude oil, gasoline, diesel and kerosene ranged from 0,126 - 3,06 µg/L. Therefore, the developed m-SPE method prior to GC-HR-ToF-MS analysis can be applied as an alternative for quantitative determination of OSCs in oily matrices.

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LIST OF ABBREVIATIONS AND ACRONYMS

µ-SPE - Solid phase microextraction AED - Atomic emission detector APS cartridge - Aminopropylsilane cartridge ASE - Accelerated solvent extraction BT - Benzothiophene CAR-PDMS - Carboxen polydimethylsiloxane CCD - Central composite design **DBT** - Dibenzothiophene **DCM** - Dichloromethane **DI-SDME** - Direct single-drop micro-extraction DLLME - Dispersive liquid-liquid microextraction Dynamic HS - Dynamic headspace ESI - Electronspray ionization FID - Flame ionization detector FLD - Flouresence detector FM-GC×GC-MS/MS -Flow modulated comprehensive two-dimensional gas chromatography-triple quadrupole mass spectrometry FPD - Flame photometric detector FTIR - Fourier transform infrared GC -HR- ToF-MS - Gas chromatography high resolution time-of-flight spectrometry GC - Gas chromatography GC×GC-AED -Comprehensive twochromatography dimensional gas atomic emission detector GC×GC-FID - Comprehensive two-dimensional gas chromatography flame ionization detector GC×GC-MS - Comprehensive two-dimensional gas chromatography mass spectrometry GC×GC-SCD comprehensive twodimensional gas chromatography sulphur chemiluminescence detector

GC×GC-ToF-MS - Comprehensive twodimensional gas chromatography time-of-flight mass spectrometry,

GC-FID - Gas chromatography flame ionization detector

GC-FPD - Gas chromatography flame photometric detector

GC-ICP-MS - Gas chromatography inductively coupled plasma mass spectrometry

GC-MS - Gas chromatography mass spectrometry

GC-MS/MS - Gas chromatography tandem mass spectrometry

GC-MS-NCI - Gas chromatography negative ion chemical ionization mass spectrometry

GC-O-MS - Gas chromatography olfactometric mass spectrometry

GC-PFPD - Gas chromatography pulse flame photometric detector

GC-Q-ToF-MS - Gas chromatography quadruple time-of-flight mass spectrometry

GC-SCD - Gas chromatography sulphur chemiluminescence detector

HF-LLME - Hollow fibre liquid-liquid microextraction

HPLC - High performance liquid chromatography

HPLC-UV - High performance liquid chromatography ultraviolet

HS-SDME - Head space single-drop microextraction

HS-SPME - Head-space solid phase microextraction

HT-GC×GC-SCD - High temperature comprehensive two-dimensional gas chromatography sulphur chemiluminescence

LC - Liquid chromatography PDMS-DVB Polymethylsiloxane LC-MS - Liquid chromatography mass divinylbenzene spectrometry RSD - Relative standard deviation LLE - Liquid-liquid extraction SBSE - Stir bar sorbtive extraction LOD - Limit of detection SCD - Sulphur chemiluminiscene detector LOQ - Limit of quantification SEM - Scanning electron microscopy SFE - Supercritical fluid extraction **MAE** - Microwave assisted extraction MALDI - Matrix assisted laser desorption SPE - Solid phase extraction ionization SPME - Solid phase microextraction, MS - Mass spectrometry **TD-GC-MS** - Thermal desorption gas **MSM** - Methylysulphonyl methane chromatography mass spectrometry mSPE - Magnetic solid phase extrcation TEM - ransmission electron microscopy N/A - Not available TLC- Thin layer chromatography NBT - Naphthobenzothiophene TP - Thiophene ND - Not detected UAE-LLME -Ultrasound assistedemulsification liquid-liquid microextraction NDBT - Naphthodibenzothiophene **NP** – Nanoparticles **UE** - Ultrasonic extraction NPLC-GC/MS Normal liquid UHPLC -Ultra-high-performance liquid phase chromatography-gas chromatography chromatography-mass spectrometry mass UV - Ultraviolet spectrometry **OSCs** - Organosulphur compounds **XRD** - X-ray powder diffraction

PDMS - Polydimethylsiloxane

CHAPTER I

INTRODUCTION

PREAMBLE

A brief background of crude oil as a fossil fuel and its origin is discussed in this chapter. Under the background section, crude oil refinery process is elaborated to produce other fuel oils. Different examples of selected OSCs are also outlined together with their negative effects. The chapter further confers on the problem statement, aims, hypothesis and the justification of the study. Lastly, the outline of the entire dissertation, which explains the content of each chapter, is summarized.

1.1. Background

Crude oil is a naturally occurring mixture of hydrocarbons and non-hydrocarbons, which makes up fossil fuel in a liquid state. Crude oil is regarded as a fossil fuel because of its origin, as it forms from fossilized buried remains of carbon rich plants and animals that lived decades of years ago [2]. The high pressures, high temperatures, microbiological activities and geological processes of the earth convert these remains into crude oil [3]. It is found in the reservoirs, cracks, narrow openings and pores of the sedimentary rocks underground; thus, it is accessible through drilling. Crude oil is sometimes found on the earth surface in the tart sands [4]. **Figure 1.1** illustrates the formation and the location of crude oil.

Crude oil is regarded as one of the essential energy sources for transportation and other industrial operations around the world. It is considered as such because, the buried crude oil is extracted from the deeper surface of the earth and refined into important products such as kerosene, diesel, gasoline and other petroleum products [5].



Figure 1.1 Illustration of fossil fuels formation and location underground.

1.2. Crude oil composition

Hydrocarbons are a major component of crude oil and they contain carbon atoms which forms different structures that can either be linear or branched [6]. The most dominant fractions in crude oil are paraffin, naphtha and aromatics (toluene and xylene as derivatives of benzene rings). Furthermore, isoalkanes and methyl cycloalkanes are the main constituents of paraffin hydrocarbons. Naphtha on the other hand, is mostly composed of five to six-membered rings, however, seven membered rings can occasionally be observed [7]. Non-hydrocarbons contain derivatives of nitrogen, sulphur, oxygen and various metal ions in trace concentration levels [8]. **Figure 1.2** represents the different compositions of crude oil.



Figure 1. 2 Molecular composition of crude oil [7].

Hydrocarbons contribute up to 60% of crude oil. The dominating paraffin structures contribute to the type of crude oil. When short chained paraffin are contained in crude oil, the crude oil is deemed light crude oil [9]. Aromatics are also unsaturated hydrocarbons that make up to 30% of crude oil [10].

1.3. Classification of crude oil

Crude oil can be classified using both physical and chemical properties. The basic physiochemical properties of crude oil are the same throughout, even though they originate from different geographic locations. However, the location determines the nature of crude oil and its composition [11]. Furthermore, crude oil samples that are extracted from the same well at different times can also differ in terms of chemical composition and physiochemical properties [7]. Fossil fuel can be divided into heavy and light fractions, where heavy oils flow slowly as compared to light oils. Therefore, physical properties such as viscosity and density are normally used to distinguish between the heavy and light crude oils. Light crude oil is less viscous, and its density is less than that of heavy crude oil. Additionally, light crude oil is less expensive to process as compared to the heavy crude oil. This is because, heavy oil contains long chains of hydrocarbons, which are difficult to distillate [12]. Another criterion used for crude oil classification is the American Petroleum Institute (API) degrees and **Table 1.1** shows the classification of crude oil according to API [13].

Crude oil	APIº
Light crude oil	<31.1°
Medium crude oil	31.1 -22.3°
Heavy crude oil	22.3 -10°
Extra heavy crude oil	>10°

Table 1.1 Classification of crude oil by API

It is well documented that sulphur is the most abundant heteroatom in crude oil with 0.03-6%wt. concentration levels [14]. Therefore, crude oil always contains certain

types of OSCs. Crude oil that has higher density generally has a low API degree. This kind of crude oil contains a high content of OSCs. The presence of these OSCs are undesirable/detrimental in crude oil products due to their harmful effects. For example, the presence of OSCs in diesel and gasoline promotes corrosion of engine parts on vehicles, leading to the production of water-containing sulphur dioxide from the exhaust as a result of the combustion process that takes place [15]. Crude oil contains less than 0.5% of sulphur, whilst above 0.5% content, it is regarded as sour crude oil [7]. Crude oil with a low sulphur content is desirable in the finished product thus, sweet crude is always preferable than the sour crude oil. Refining of sweet crude oil is easier compared to sour crude oil, since less sulphur results in less damages in the refineries [16]. The OSCs include mercaptans/thiols, thiophenes, just to name the few. Inorganic sulphur species are sulphates, thiosulphate, sulphides and sulphites [17]. For this study, the focus was on thiophene and its derivatives and **Table 1.2** shows selected structural examples of typical OSCs.

Thiophenes are one group of OSCs contained in crude oil. These are a fivemembered electron-rich heteroatomic OSCs that are liquid at room temperature. The thiophene compounds are toxic and flammable. They are insoluble in water but soluble in organic solvents such as alcohol and ether [18]. Polycyclic aromatic heteroatomic sulphur compounds (PASHs) include dibenzothiophenes (DBT), benzothiophenes (BT) and Naphthobenzothiophenes (NBT). These PASHs are the most dominating and challenging OSCs to remove during the process of refining, due to their aromatic ring character [19]. Thiols are also knowns as mercaptans and they are analogous to alcohols. They are identified by the –SH group and characterized with a pungent odour [20].

The concentration levels of OSCs in crude oil varies in relation to the geographical location of the crude oil [21]. For example, Nigeria contains between 0.04 - 0.25 w/w% and Egypt, which is another African country contains 0.04-4.19 w/w% of OSCs in crude oil. European countries such as Norway and Denmark contain between 0.04-0.67 w/w% and 0.2- 0.25 w/w % of sulphur, respectively [22].

Table 1. 2 Examples of selected OSCs [23]

Organosulphur group	Molecular formula	Organosulphur example	Skeletal structure
Thiol/Mercaptan	R-SH	Thiophenol	SH
		Propanethiol	SH
Disulphide	R-S-S-R	Dimethyl disulphide	s s
Thiophene (Th)	C4H4S	3-methyl thiophene	S S S S S S S S S S S S S S S S S S S
		2-ethyl thiophene	
Benzothiophene (BT)	C ₈ H ₆ S	2,3-dimethyl benzothiophene	



1.4. Problem statement

Organosulphur compounds (OSCs) are a major contributor to air pollution. During the refining process of crude oil, OSCs emit oxides of sulphur (SOx), which contributes to the formation of acid rain, thereby damaging the environment and the surface of building structures [3]. Sulphur emissions can also interact with ozone and other gases to produce sulphur containing smog that can be harmful to human and animal health [8]. Furthermore, the presence of OSCs in crude oil can also lead to the deactivation of the catalyst during the refining processes. The quality of crude oil is the determinant factor in terms of price differentials. Stringent sulphur regulations and specifications have been adopted in various countries to minimize its toxicity to the environment. The United States of America (USA) and European Union (EU) for example, reduced the concentration of sulphur in diesel from 500 to 15 mgL⁻¹ and 50 to 10 mgL⁻¹respectively [10].

Therefore, the knowledge of OSCs structures in crude oil is a key factor in the development of effective sulphur removal processes. The development of novel crudeoil desulphurisation catalysts requires detailed knowledge of sulphur-containing compounds present in crude oil and intermediate products. Many of these OSCs are volatile in nature and some are in trace concentration levels in a complex crude oil matrix. The nature of complex crude oil matrix makes it challenging for qualitative and quantitative determination of OSCs [11–13].

Different chromatographic methods have been employed for the determination of OSCs compounds in crude oil. Most of these techniques involves sample pre-treatment prior to analysis. Sample pre-treatment is necessary in order to pre-concentrate and/or to pre-clean the crude oil sample because of its complexity. Liquid-liquid extraction (LLE), solid phase extraction (SPE), solid phase micro extraction (SPME), hollow fiber liquid phase extraction (HF-LPME) and dispersive liquid-liquid micro extraction (DLLME) are some of the sample pre-treatment methods that have been reported for the analysis of OSCs in crude oil [24]. Some of these techniques use organic solvents that are halogenated in large amounts. These solvents can be toxic and are not easy to handle, thus can in turn be harmful to the handler. Furthermore, the sample required for some of the techniques should be in large quantities and this is not in-line with green chemistry [25].

1.5. Aim and objectives

The main aim of the study was to develop a rapid and selective analytical method for qualitative and quantitative analysis of organosulphur compounds (OSCs) in selected fuel oil samples using GCTOFMS.

The specific objectives of this project were to develop:

- Dilution pre-treatment method prior to qualitative determination of organosulphur compounds in various crude oil samples using GCTOFMS.
- Magnetic solid phase extraction (m-SPE) followed by GC-TOFMS for semiquantitative analysis of organosulphur compounds in various fuels.
 - Magnetic gold nano-composites were synthesised and characterised using various techniques such as TEM, SEM-EDX, P-XRD, UV-Vis and FT-IR.
 - The exploration of various parameters (adsorbent mass, eluent type and volume, extraction time and pH) that affect the extraction efficiency of the proposed m-SPE was performed by using multivariate optimization methods.
 - The analytical merit (performance) of the proposed m-SPE method was compared with literature reported pre-concentration procedures.
 - The optimum m-SPE sample preparation procedure was applied in real oil samples for the pre-concentration/extraction of organosulphur compounds.

1.6. Justification of the study

Chromatographic techniques have been used for the determination of OSCs in various samples. These chromatographic techniques include a high performance liquid chromatography (HPLC) [26], Gas chromatography (GC) [27] and a comprehensive two dimensional gas chromatography (GC×GC) [28] amongst others. A sample can be analysed directly using chromatography with any minimal sample pre-treatment [27]. Minimal sample pre-treatment can be a dilution of the sample using organic solvents such as heptane [29], dichloromethane (DCM) [30] *etc.* prior to analysis. HPLC was

used by Silveira and co-workers to analyse OSCs in asphalt cement. Toluene and methanol were used as solvents for diluting the samples. This method achieved an accuracy of between 83.85 % and 110.28 %

However, OSCs are found in low concentrations in fuels [14], thus sample preparation or pre-treatment is important prior to analyses in order to improve detection limits [31]. Due to the complexity of a crude oil sample, different sample pre-treatment techniques have been employed for the analysis of OSCs. These techniques include the traditional methods such as LLE [25] and SPE [35] amongst other techniques. The OSCs such as benzothiophene and dibenzothiophenes have been determined using the former technique. From their findings, the extraction of OSCs was between 0.10-250 µg mL⁻¹ and a recovery of 77-91% was achieved [36]. This technique also uses solvents, which can be toxic, thus the latter was employed as a solvent less technique. Silveira et al [37] used aminopropyl cartridges as adsorbents and a better recovery of 65.37 to 114.85% was achieved. It also yielded a better extraction of OSCs (0.1-3.5 mgL⁻¹) [37]. The sorbent used was commercially available it can be relatively costly. Other researchers synthesized 8- hydroxyquinoline silica gel impregnated with palladium chloride as an adsorbent which was also expensive. A recovery between 81.5-92.1% was obtained using this technique [38]. The SPE is rapid, and this method was shown to have a high enrichment factor and a high recovery. This has made SPE to be a popular strategy for effective and selective pre-concentration of analytes [39]. Magnetic nano composite adsorbents have received a lot of focus because of their unique structure, properties, high adsorption ability, and reusability [40]. During literature compilation, no information was found in relation to use of the nano composite for OSCs analysis.

Thus, this study seeks to use GC-TOFMS for qualitative and semi quantitative analysis of OSCs in oily matrices after sample pre-treatment. Four target OSCs were chosen for the study due to time limitations. This chromatographic technique has received a lot of attention from researchers because it provides additional separation of overlapping species [41] as well as superior resolution with high sensitivity [42].

1.7. Hypothesis

Dilution (direct analysis) and magnetic solid-phase extraction (indirect analysis) can be used as sample pre-treatment methods for qualitative and quantitative determination, respectively of OSCs in fuel oils using a comprehensive GC-HR-ToF-MS. The gold magnetic nanoparticles can be applied as selective adsorbents for OSCs during m-SPE and the magnetic character will enhance easy separation of adsorbent from the bulk of the solution.

1.8. Research questions

This study seeks to answer the following research questions:

- Does the use of dichloromethane as a solvent aid in separation of OSCs peaks when using a gas chromatography coupled to high resolution time-of-flight mass spectrometry (GC-TOFMS)?
- Is gold coated magnetite selective towards OSCs on the selected fuel samples?
- Does the developed magnetic solid phase extraction method of OSCs applicable to real fuel oil samples?
- How will the figure of merits of this study compare to the previously done studies?

1.9. Dissertation outline

Chapter I (Introduction)

This chapter deals with background information about crude oil. It further explains the origin of crude oil and its composition. An introduction to organosulphur compounds that are present in crude oil is presented. The hypothesis, objectives, problem statement, research questions and the justification of the study are outlined.

Chapter II (Literature review)

The literature on qualitative and quantitative analysis of OSCs in crude oil is discussed extensively in this chapter. Different chromatographic techniques that are associated with the analysis of OSCs in crude oil are highlighted with detailed information on how the analysis were conducted. Sample pre-treatment methods that are used for pre-concentration and sample clean up prior to chromatographic analysis are evaluated. Furthermore, the direct methods of analysis that include dilution are detailed in this chapter. It is worthy to indicate that this chapter has been published as a review article [43].

Chapter III (Methodology)

All the procedures and methodologies that were followed in order to generate data for this study are well defined in this chapter. This includes all general laboratory techniques that were used for qualitative and quantitative analysis of OSCs in crude oil and the characterization techniques for the confirmation of the synthesised adsorbents for m-SPE experiments are described. A detailed description of multivariate optimization methods is also expressed in this chapter.

Chapter IV (Results and discussion)

The qualitative data for the analysis of OSCs in crude oil is presented in this chapter. The discussion of the data is also focussed on. The optimization parameters and results are also presented in this chapter. The precision of the method is also compared against the already published literature reports. Furthermore, this chapter also focuses on the quantitative results and discussions including the figures of merits.

Chapter V (Conclusion and future recommendations)

Overall findings in relation to the aim of the study are summarized in this chapter. Furthermore, recommendations and future work are also presented in this chapter.

CHAPTER II

LITERATURE REVIEW

PREAMBLE

In this chapter, chromatographic analytical techniques that are mostly reported for both qualitative and quantitative determination of OSCs in various matrices are described based on direct (analysis without sample preparation) and indirect (analysis after sample preparation) analysis. The merits and possible limitations associated with direct and indirect analysis are also discussed. Additionally, the analytical performances of the various published sample preparation methods are evaluated in terms of accuracy, the limit of detection, precision, etc. Lastly, this chapter highlights current gaps in chromatographic determination of OSCs and possible future developments.

2.1. Introduction

Organosulphur compounds (OSCs) have a carbon-sulphur bond, exist in different chemical structures and are present in various matrices such as vegetables [44], wine [45], crude oil [46], coal [47], etc. It has to be noted that, OSCs are naturally occurring thus their presence is unavoidable [48]. Some of these sulphur compounds are even found in living organisms as amino acids (e.g. methionine, glutathione and thiamine) [49]. The OSCs are categorised into different classes depending on the functional group of the sulphur atom. The commonly documented OSCs include thiols (mercaptans), sulphides, disulphides, thiophenes, benzothiophenes and dibenzothiophenes [7].

Sulphur compounds such as sulphoxides and sulphones are mostly used as manufacturing products in pharmaceutical industries and other compounds like methylsulphonylmethane (MSM) are supplements that are present in foods for the biological maintenance of hair, skin and nails [50]. The most dominating OSCs present in crushed garlic cloves are sulphoxides like alliin and are good anti-carcinogenic and anti-oxidant reagents [51]–[53]. An investigation by Hall and co-workers was conducted in relation to the presence of OSC in garlic and it was concluded that, OSCs can also reduce inflammation and oxidative stress [54]. It must be noted that, onion is another alliin containing vegetable, with similar biological effects as garlic. Volatile OSCs (diallyl disulphide, allyl methyl trisulphide and dipropyl disulphide) contained in garlic and onion

vegetables are mostly responsible for the anti-inflammatory and extra ordinary antimicrobial activities [55].

Alcohol beverages such as whisky, wine and beer also contain OSCs, which are mostly responsible for flavour improvement. However, the concentration levels of these compounds vary from one beverage sample to another. For example, fusel alcohol and whisky contains μ g L⁻¹ levels of OSCs[56]. The OSCs like thiols, sulphides and disulphides are found in low concentrations in wine and are mainly accountable for the distinct aroma of the wine [57].

The presence of OSCs can either be essential or toxic, depending on their concentration levels, chemical structures and the host sample matrices [51]. OSCs present in environmental samples (water and sediments) and in fossil fuels (coal and crude oil) are mainly detrimental. Gimeno and co-workers reported the presence of polycyclic aromatic sulphur heterocycles (PASHs) in seawater and sediment samples and their potential genetic mutation and cancerous activities [26]. Additionally, it was reported that the commonly found PASH in environmental samples such as spills and discharges (regulated and unregulated) is thiophene. The latter showed some toxicological effects, which include cancer and genetic mutations. [58]. These thiophenic sulphur compounds are abundant in fossil fuels such as crude oil. Burning of the sulphur containing fossil fuels results in the emission of toxic sulphur gases (SO₂ and SO₃) to the atmosphere. These gases are major contributors to the formation of acid rain, which is harmful to plantation, living organisms and building infrastructure [59]. Furthermore, the produced SO₂ can interact with the ozone, results in formation of harmful smog and cause negative respiratory effects to humans [60]. It is also documented that only 1 ppm of SO₃ can lead to coughing and choking in human beings, thereby causing chronic diseases such as bronchitis [61]. Since these fossil fuels are naturally occurring, the amount of OSCs differs depending on geographical location. For example, Australia, Denmark, and Nigeria's fossil fuels contain less than 0.26 %(w/w) of the OSCs and other countries like Cuba contain as high as 7.03 %(w/w) of OSCs [19]. Due to the toxicity of these compounds, different countries adapted stringent regulation conditions to minimize the harmful impact of these compounds to the environment. For example, USA allows the concentration of OSCs to be between 15000 to 500000 μ g L⁻¹, while EU is only allowing 1000 to 5000 μ g L⁻¹ of OSCs in diesel oils [62].

It is therefore crucial to monitor the presence of OSCs and their concentration levels in different matrices. In petroleum industries, the presence of OSCs can lead to the deactivation of the catalyst during refining processes. The knowledge of OSC structures is a key in the development of effective desulphurization processes in various industries such as pharmaceutical, agriculture, food and petroleum industries [63]. The development of novel desulphurisation processes requires detailed knowledge of OSCs structure and concentration level.

2.2. Determination of OSCs in different matrices

High performance liquid chromatography (HPLC) [64], gas chromatography (GC) [65] and thin-layer chromatography (TLC) [66] are some of the chromatographic techniques that have been previously reported by various researchers for OSCs determination in various matrices. The TLC relies on stationary phase that is coated on glass or plastic plate. Samples are mounted on the plate in the presence of a liquid mobile phase which is a solvent or a mixture of solvents and placed on a chamber. The sample migrates through the plate on the basis of the analytes' physiochemical properties which determines the interaction with the mobile and stationary phase [67]. Unlike TLC, HPLC uses a column, and the analytes of interest are separated in the column. The packing of the column considers different properties such as the pore sizes, the size of the particle and the functional groups. The normal phase and the reverse phase liquid chromatography are the most popular types of chromatography for OSCs determination [68]. Gas chromatography (GC) is another separation technique that has been used for the determination of volatile OSC in various complex matrices. It must be noted that GC operates similarly to the HPLC. However, GC techniques use inert gasses as mobile phase (helium, nitrogen and hydrogen). This technique offers an opportunity of a wide range of analysis of compounds in different matrices, but more volatile analytes are recommended for efficient chromatographic separation. The stationary liquid phase in GC column is volatized using the temperature gradient. Therefore, easily decomposed organic compounds are preferably analysed by other chromatographic techniques [69], [70].

In order to obtain a more efficient separation, researchers have explored the use of a comprehensive two dimensional chromatography (GC×GC) [41]. The latter is a major analytical tool that overcomes drawbacks that were found in other chromatographic techniques. This technique can overcome challenges such as the presence of OSCs in low concentrations and composition of the OSC in complex matrices [71]. This is because, GC×GC uses dual columns which are coupled to a modulator. The separation on both columns takes place in gaseous phase. The first column is wider and longer than the second column. Two independent steps of separations were combined into one when using this technique. Some of the fraction in the first column go to the second column where they undergo further separation [72]. The GC separation is conducted by using different of types stationary phase combinations [41], [73]. Therefore, this chapter discusses literature reported methods applied for OSCs determination using direct (without sample preparation) and indirect (with sample preparation methods followed by analysis) prior to chromatographic analysis, as shown in Figure 2.1.



Figure 2. 1 Direct and indirect methods used for chromatographic analysis of OSCs

2.2.1. Direct chromatographic analysis of OSCs in various matrices

In this technique, a sample is analysed directly without undergoing any sample pretreatment prior to analysis. Direct analysis is fast and simple but is often more suitable for highly concentrated analytes. This analytical strategy does not involve many steps, thus the samples are normally less contaminated [74]. Additionally, direct analysis minimizes the loss of target analytes, which may occur during the sample pre-treatment stages. However, this analysis is only possible when the concentration levels of the OSCs in a particular matrix are higher than the detectors detection limit of the suitable chromatographic technique and when the matrix is less complex [75]. Solvents such as dichloromethane (DCM) [25], n- hexane [76], xylene, toluene [77], 2- propanol and octane [78] are commonly used for dilution of highly viscous samples prior to chromatographic analysis of OSCs. These solvents should be compatible with the proposed chromatographic technique and be able to dissolve the sample and the analyte without affecting the composition of the sample and the chemical structures of the target analytes [79]. It has to be noted that, complex samples like crude oil should be diluted with organic solvents that have low boiling points and high surface tension and the separation between the sample and the solvent should be fast and have chemical stability [80]. Although dilution is a simple technique that is less laborious, it has its own challenges. For example, some of the organic solvents used during sample dilution are difficult to handle, due to their vapour pressure and low viscosity [81]. Additionally, the concentration of the target analyte can increase if using contaminated solvents. It has been reported that, some organic solvents are toxic and hazardous, thereby, require care handling [81]. However, there are samples such as crude oil that poses challenges with regards to solubility [82].

A summary of different direct chromatographic analytical methods for the determination of OSCs in various matrices is presented in Table 2.1. It must be noted that, there was either dilution or no sample pre-treatment prior to analysis. The dilution sample pre-treatment was conducted by using organic solvents. This table clearly outlines that GC techniques were the most widely used techniques focusing on both the two dimensional and one-dimensional chromatography from the gathered information. This may be due to advantages of GC techniques towards more volatile compounds and complex samples such as crude oil. It can be further deduced from Table 2.1 that dibenzothiophene (DBT) was the most determined analyte and

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naphthodibenzothiophenes (NDBT) was the least determined analyte. Additionally, cyclic OSCs were the most target analytes compared to the others such as sulphoxides and thiols. The favour in DBT analysis may be due to the fact that direct analysis was mostly investigated on petroleum samples, where DBT causes challenges and is difficult to remove [21]. Therefore, samples that contain cyclic OSCs were less favoured in direct analysis, due to the challenges that these compounds bring during GC analysis. Cyclic organosulphur compound, alkylbenzothiophene was the analyte of interest during the study by Zhu et al. in a diesel sample. This compound was directly determined quantitatively without any sample pre-treatment method. The diesel sample was injected into a GC hyphenated to a quadruple time-of-flight mass spectrometry (GC-Q-ToF-MS) using a split/split less injector. This method resulted in a good correlation coefficient that is greater than 0.9920. The method validation included precision and repeatability. The former was calculated using the relative standard deviation (RSD) and was found to be between 1.1-3.4%. The accuracy was 93.1-107.8% for all the dibenzothiophene compounds determined [27]. HPLC coupled to an atmospheric pressure photoionization tandem mass spectrometry for the determination of OSCs in asphalt cement was reported. During this study, Silveira and co-workers used toluene fractions and methanol for dilution of asphalt cement and the HPLC technique detected 14 OSCs (sulphoxides, sulphones and dibenzothiophene) with accuracy between 83.85 and 110,28% for some of those compounds [31]. The DCM solvent was also used by Wilson et al during the qualitative characterization of polycyclic aromatic OSCs by normal phase liquid chromatography [83]. Dichloromethane was used to dilute coal tar, diesel particulate and petroleum extracts. The precision, LODs, LOQs and the recoveries were not reported [83]. Gas chromatography was coupled with a high-resolution mass spectrometry for the determination of alkylbenzothiophenes (alkyl-BT) and alkyl-dibenzothiophenes (alkyl-DBT) in light cycle oil and straight run gas oils. The results were compared to those of GC-SCD and it was reported that the distribution of thiophenic sulphur type was 10% and that of non-thiophenic was 90% in straight run oils. Furthermore, 1.19% of sulphur was contained in gas oil [84]. The percentage of OSCs in crude oil from three geographical locations was characterized. Crude oil that originated from Italy was found to have between 2.1% and 7.1% level of OSCs. The lowest quantity of OSCs was obtained from crude oil that originated in Africa with 0.2%. Oil from Asia had 1.2% and 3.8% of OSC levels. During this study, the limit of quantification of this method was obtained as 2.4 -5 ppb [33]. Guy and co-workers for the identification of benzothiophene, dibenzothiophene and naphthothiophene in light cycle oil used atomic emission detector (AED). The distribution of these OSCs and their retention times were determined by directly injecting the sample at a split ration of 59:1 in GC. The substituted benzothiophene was found to be in majority with 59.3% and 40.7% being shared between dibenzothiophene and naphthothiophene [85]. The one dimensional chromatography was unable to separate and quantify some of the OSCs contained in complex crude oil samples without sample pre-treatment [86]. Therefore, the application of GC×GC, (which is known as a powerful tool for quantitative and qualitative analysis of OSC) was reported. For example, Avila and co-workers reported the use of GC×GC-ToF-MS for both qualitative and semi quantitative determination of OSCs in crude oil. The latter was diluted with DCM solvent prior to OSCs analysis and *n*- tetracosane d₅₀ was used as an internal standard. The chromatographic results showed the presence of sulphides, disulphides, and thiols with a total concentration of 1.372 μ g L⁻¹. However, researchers did not report the validation of the analytical method [87].

Table 2.1 further indicates that direct sample injection without minimal sample pretreatment is not often used compared to minimal sample pre-treatment such as dilution and DCM was the mostly used diluent when minimal sample pre-treatment was applied. This can be attributed to the nature of this diluent as it is a polar solvent and miscible with most solvents. Although it is highly volatile which makes it hazardous when inhaled, it is less toxic when compared to other halogen containing solvents [88]. When this diluent was used, a good linear response was also achieved [31]. There are other diluents that were reported, such as toluene [33], isooctane [34] that also yielded good responses for the analysis of these OSCs.

Matrix	OSCs	Detection Technique	Diluent	LOD (µg/g)	Accuracy (%)	Precision (%RSD)	Linear Range (µg/g)	Correlation coefficient R ²	Ref
Asphalt cement	Sulfoxide, BT, DBT, NBT	UHPLC-MS/MS	DCM	0.28-5.23	83.85- 110.28	1.04	50 - 1500	>0.99	[31]
Coal	DBT	NPLC-GC/MS	DCM/Toluene	ND	ND	ND	ND	ND	[83]
Coal tar	DBT	NPLC-GC/MS	DCM/Hexane	ND	ND	ND	ND	ND	[89]
Crude oil	DBT isomers	GC – MS/MS	Toluene	0.7	ND	0.68-2.01	ND	ND	[33]
Crude oil	BT, DBT	GC – MS/MS	Isooctane	0.0099	83.8-103.0	0.2 – 5.6	0,020- 9.99	0.9999	[34]
Crude oil	DBT	GC×GC-ToF-MS	DCM	ND	ND	ND	ND	ND	[28]
Crude oil	Thiols, sulphides, TP, BT, DBT, NBT	GC×GC-SCD	N/A	ND	90-102	6.02	ND	ND	[90]
Diesel	DBT	GC-Q-ToF-MS	N/A	3000	93.1-107.8	1.1 – 3.4	20-2000	>0.9920	[27]
Gas Condensate	Thiols, BT, DBT	GC-SCD	Isooctane	ND	ND	5-6	ND	ND	[91]
Gas oil	BT, DBT	GC-SCD	Toluene	ND	<100	ND	ND	0.999	[84]
Heavy crude oil	DBT	GC-SCD	<i>n-</i> Hexane/DCM	ND	ND	ND	ND	ND	[92]

Table 2. 1 Direct analysis of OSCs in different matrices using chromatographic techniques

Heavy petroleum	TP, BT, DBT, NDBT	HT-GC×GC-SCD	Toluene	ND	ND	ND	ND	>0.9985	[93]
Heavy gas oil	TP, BT, DBT, NBT	FM-GC×GC- MS/MS	DCM	6-90	ND	7-17	1-100	>0.9975	[30]
Light cycle oil	BT, DBT	GC-SCD	n-Octane	ND	ND	ND	ND	ND	[94]
Petroleum samples	TP, BT, DBT, NBT	GC×GC-TOF-MS	Tetracosane/ pyrene	ND	ND	ND	ND	ND	[87]
Petroleum products	TP, BT, DBT	GC-ICP-MS	<i>n</i> -Hexane	7	ND	ND	ND	ND	[95]
Petrochemical samples	BT, DBT, NBT	GC×GC-AED/MS	N/A	ND	ND	ND	ND	ND	[96]
Shale oil	TP, BT, DBT	GC×GC-FPD	DCM	20	ND	10 - 35	0.025-1	>0.994	[97]

ND = Not determined, N/A = Not available, OSCs = organosulphur compounds, TP = thiophene, BT = benzothiophene, DBT = dibenzothiophene, NBT = naphthobenzothiophene, NDBT = naphthobenzothiophene, NDET = naphthobenzothiophene, NDET = naphthobenzothiophene, GCxGC-SCD = comprehensive two dimensional gas chromatography sulphur chemiluminescence, FM-GCxGC-MS/MS = flow modulated comprehensive two dimensional gas chromatography inductively coupled plasma mass spectrometry, GCxGC-AED = comprehensive two dimensional gas chromatography atomic emission detector

2.2.2. Indirect chromatographic analysis of OSCs in various matrices

Some samples are complex and contain low concentration levels of OSCs, thereby causing various challenges during chromatographic analysis. To circumvent these challenges, sample preparation techniques have been introduced [24]. Depending on the complexity of the sample, sample preparation could involve a couple of steps. The complexity of the sample may include, the concentration levels of the OSCs in the matrix of interest, sample state of the matter (gas, liquid or solid) and the physiochemical properties of target OSCs [100]. Furthermore, variables such as the limit of detection (LOD), limit of quantification (LOQ) linearity, correlation efficient and accuracy are also affected by the choice of the sample preparation methods used [69]. In the current literature review, different sample preparation methods such as solid phase extraction (SPE), liquid-liquid extraction (LAE) and other {supercritical fluid extraction (SFE) and ultrasound assisted extraction (UAE)} are extensively discussed and evaluated according to their performance for chromatographic determination of OSCs in various samples.

2.2.2.1. Liquid-Liquid extraction of OSCs in various matrices

Liquid-liquid extraction (LLE) is one of the traditional sample pre-treatment methods that have been employed with the view of increasing target analyte sensitivity. The LLE is achieved by separating the target analytes from the major sample matrix and other interfering species through partitioning between two immiscible liquids [101]. The LLE was improved to a miniaturized technique called Liquid liquid-microextraction (LLME) technique that is harmonious with GC and liquid chromatography [102]. The LLE procedure incorporates three steps, which are sample clean-up, extraction of analyte and pre-concentration [103]. This method was employed as a sample pretreatment procedure for the analysis of OSCs in wine samples. The extraction recoveries (91.99-125.87%) were good with effective separation. However, the involvement of numerous steps and large quantities of reagents make LLE time consuming, tedious, expensive, harmful to the user and might introduce crosscontamination [102]. Therefore, flotation assistance was applied together with the LLE method for the extraction and determination of OSCs in water samples. During this study, acetone was used as a homogeneous solvent and toluene was used as an extraction solvent. It was concluded that this method was efficient and the linearity was in the range of 0.25-200 μ g L⁻¹ with the LODs in the range of 0.05-0.4 μ g L⁻¹ [1].
Liquid-liquid micro extraction (LLME) uses microliter quantities of solvent for preconcentration of analyte [104]. This technique was introduced to solve challenges that were experienced in LLE and it is simple, rapid and inexpensive [105]. There are three categories of LLME namely single drop micro extraction (SDME), dispersive liquidliquid micro extraction (DLLME) and hollow-fibre liquid phase micro extraction (HF-LPME) [106]. Xiong and Hu compared DLLME and HF-LLME for the analysis of OSCs in environmental and beverage samples. During this study, GC was coupled to FPD and the relative standard deviations of 8.5-13.7% were reported for DLLME. The LOD of HF-LLME were found to be 1.16 μ g L⁻¹, with the precision of 3.4%. Therefore, it was deduced that the extraction efficiency of HF-LLME was more effective than that of DLLME because it has less extraction time and higher extraction efficiency. Carbon tetrachloride was employed as a solvent for DLLME and o-xylene was used as the extraction solvent for HF-LLME [107]. The LLME technique was applied for the determination of OSCs in crude oil, with accuracy between 77-91% [103]. Liquid-liquid extraction methods were compatible for samples that are in liquid phase and the usage of the commercially available fibre increases the cost of HF-LLME methods [108].

Table 2.2 below summarizes different chromatographic analytical techniques employed by researchers for the analysis of OSCs in different sample matrices. The LLE technique demonstrates efficiency on samples that are in liquid phase or aqueous phase. Only one-dimensional gas chromatography was used with LLE as a sample pre-treatment technique. The GC mostly was coupled to a flame ionization detector with DLLE as a sample pre-treatment technique. Acetonitrile and chlorobenzene were used as solvents during a study for the analysis of OSCs in aqueous samples with DLLME employed as a sample pre-concentration technique. The latter was employed as an extraction solvent and the formers was used as dispersive solvent [109]. Lastly, crude oil was the only sample matrix where sulphur specific detector was coupled to GC. However, FPD is not regarded as a specific detector because it is sensitive to both phosphorus and sulphur compounds.

In conclusion, **Table 2.2** shows that the DLLME was the commonly used LLE technique from the extracted literature reports. This technique uses less volume (micro-litres) of solvent, it is simple to operate, it is less expensive and also it a rapid method [102] with high recovery as indicated in **Table 2.2**. The extraction time for this method is the lowest compared to the other LLE techniques, thus it is regarded as rapid. These factors can be the motivation behind the common use of this extraction

method compared to the other LLE methods.

Sample	Detection Technique	OSC	LLE	LOD (µg L ⁻¹)	Linear Range (µg L ⁻¹)	Extraction Time (min)	R ²	Recoveries %	Precisio n (%RSD)	Ref
Aqueous sample	GC-FID	Sulphides, thiols	DLLME	0.1 – 0.9	1-200	ND	>0.9904	56.4 – 76.8	<10	[109]
		Sulphides	DI - SDME	10.7 -43	100 -1000	2	0.9986	ND	18.3	
Crude oil	GC-SCD	DBT, BT	LLME	1- 50	100 – 250000 –	5- 60	0.985	77 -91	4.5 -13.3	[103]

 Table 2. 2 Indirect analysis of OSCs in different matrices using LLE followed by chromatographic techniques

ND = Not determined, OSCs = organosulphur compounds, GC-FID = gas chromatography flame ionization detector, , GC-SCD = gas chromatography sulphur chemiluminescence detector,

2.2.2.2. Solid phase extraction of OSCs in various matrices

Solid phase extraction (SPE) is one powerful pre-treatment technique that is widely used for isolation, concentration, clean up, and medium exchange procedures. This extraction method uses cartridges which are filled with the matrix compound and the compounds are extracted in relation to their interaction (ion exchange, polarity or nonpolarity interaction and size exclusion) [35] with the stationary phase. Solid phase extraction is a conventional solvent free pre-treatment method that combines both sampling and pre-concentration of analyte in one step [113]. In this technique, the analyte is distributed between the liquid phase and the stationary phase. The stationary phase is the adsorbent, and they exist in different forms. It can be silicabased, modified with NH₂, carbon based sorbents, polymer based and natural sorbents such as cotton [114]. Some of these adsorbents such as the silica-based one are not stable at extreme pH values. The SPE technique has its drawbacks, which includes poor recoveries because of the interaction between the matrix and sorbent, sorbent bed clogging when disks and cartridges are used as sorbents [106]. To minimize some of the drawbacks of SPE, a miniaturized SPE technique was derived, and it was called SPME. This technique has many advantages compared to the conventional SPE. These advantages include high consistency, rapidness, simplicity and compatible with most analyte separation [106]. The SPME technique can be divided into three modes: direct-immersion solid phase micro-extraction (DI-SPME), Head space solid phase micro-extraction (HS-SPME) and the membrane protected [115]

Table 2.3 summarizes SPE and SPME methods applied with different adsorbents for OSCs extraction prior to chromatographic analysis. A reliable quantification of polycyclic aromatic sulphur heterocycles was conducted using GC coupled to a mass selective detector (MSD) and atomic emission detector (AED) on a fossil fuel related sample. During this study, SPE with aminopropylsilane cartridges was used as a clean-up procedure for the removal of polar components in oil. Information such as the molecular weight of thiophenes and dibenzothiophene isomers were reported. The repeatability, linear range and LOD were some of the factors that were not reported [116]. Rudzinski and Rai used tandem mass spectrometry during their study for the detection of polyaromatic sulphur heterocycles in crude oil. In this report, complex crude oil mixtures were separated by using distillation, alumina, ligand exchange chromatography and SPE with LC-NH₂

cartridges. The MS detection confirmed the presence of dibenzothiophene and benzonaphthiophene derivatives in the Arabian crude oil. Post column addition of tropylium cations was performed and the retention time for the polycyclic aromatic heterocycles was 3,28 and 3,51 minutes for dibenzothiophene isomers [73]. Solid phase extraction cartridges are commercially available, thus they can be expensive to obtain [117]. The development of more selective and more easy to use adsorbents has resulted in a simplified procedure [118]. 8-Hydroxyguinoline silica gel impregnated with palladium chloride was synthesized by Yang and co-workers. The silica gel was used as an adsorbent during the determination of polycyclic aromatic sulphur aromatics in sea crude oils. The SPE pre-treatment technique was coupled with GC-MS and it was promising for quantitative analysis of polycyclic aromatic heterocyclic sulphur compounds found in low concentrations. This is because it yielded good accuracy from 81,5 to 92,1% for these compounds. Furthermore, the LOD of benzothiophene was obtained at 0.2777 μ g L⁻¹, for dibenzothiophene was at 0.597 μ g L^{-1} and finally that for naphthobenzothiophene was 0,597 µg L^{-1} [38]. Da silveira and co-workers employed aminopropyl as a stationary phase for clean up in the analysis of OSCs in petroleum asphalt cement using ultra HPLC. The use of the commercially available aminopropyl cartridges eliminated the modification of stationary phase with metals. This clean-up procedure resulted in identification of at least seven OSCs. The accuracy for these compounds were in the range of 65.37-114.87%. The relative standard deviation was less than 10.11%, this was for all the analytes [119]. The use of aminopropyl was also reported by Anderson and was modified with other metals as a clean-up method during the analysis of OSCs using liquid chromatography [120].

Solid phase micro-extraction is a solvent free technique that involves direct extraction of the analyte of interest. The method uses a diameter silica coated fibre which has a small diameter as the stationary phase [121]. The fibre is used as an interface in chromatography and is used to isolate and concentrate the solute. The analyte, sample and extraction medium are partitioned [122]. When equilibrium is reached, the absorbed solute is then transferred to the chromatographic unit for analysis. This method is less tedious and simple when compared to other sample pretreatment techniques such as liquid-liquid extraction. The volume of the samples should not be in large amount and also it can be fully automated [115]. Some fibres used for the SPME technique are not sensitive to other mixtures of volatile compounds. This was evident when Pelusio and co-workers analysed OSCs in black and white truffle aroma [123].

The SPME is the most used technique compared to all techniques as it can be seen in **Table 2.3**. The high number of publications on this technique can be attributed to the previously mentioned reasons, which includes the solvent free advantage. Furthermore, different modes of this technique are available. **Table 2.3** further highlights that the most used adsorbent is the commercially available adsorbent compared to the synthesized. The commercially available adsorbent can be relatively costly. Hence, there's a need for environmentally friendly and recyclable adsorbent for the analysis of these OSC in different samples is the SPE.

Sample	Detection Technique	Adsorbent	OSC	SPE type	Extraction time (min)	LOD µg L ⁻¹	Linear range µg L ⁻¹	Correlation coefficient	Accuracy %	RSD %	Ref
Asphalt	LC	$SiO_2C_3-NH_2$,	Sulfoxides, BT,	SPE	ND	ND	100-	0.991-	65.37-	6.03 -	[119]
cement		SiO ₂ /C ₃ -CN	DBT, NDBT				3500	0.997	114.58	10.11	
Crude oil	GC-MS	8-HQ-Pd-silica	BT, DBT, NDBT	µ-SPE	2	0.193-	10 -	0.9894–	81.5-92.1	3.23	[38]
						0.597	1000	0.994		_	
										4.20	
Diesel	GC×GC-	Pd-Al ₂ O ₃	Sulphides, BT,	SPE	ND	ND	ND	ND	ND	ND	[128]
	ToF-MS		DBT								
Fossil fuel	GC-	APS cartridge	NDBT, DBT	On-	ND	ND	ND	ND	ND	ND	[116]
	MS/AED			line							
				SPE							
Gas	GC-MS	CAR-/PDMS	Sulphide	SPME	20	1x10 ⁻⁶ –	0.1-	0.992-	ND	2-12	[132]
matrices						4.7x10 ⁻	100	1.000			
						4					
Gas	GC-FID	CAR-PDMS	Thiol, sulphides	SPME	5	ND	ND	ND	ND	ND	[133]
samples											

Table 2. 3 Indirect analysis of OSCs in different matrices using SPE followed by chromatographic techniques

Lichens	GC-	Glass Cartridge	DBT	µ-SPE	N/D	<1	<2500	0.99	70 -130	16	[134]
	MS/ToF-										
	MS										

ND = Not determined, N/A = Not available, GC-MS = gas chromatography mass spectrometry, LC = liquid chromatography, BT = benzothiophene, DBT = dibenzothiophene, NDBT = naphthodibenzothiophene, SPME = solid phase microextraction, SPE = solid phase extraction, HS-SPME = head-space solid phase microextraction, μ -SPE = solid phase microextraction, APS cartridge = aminopropylsilane cartridge, GC-FID = gas chromatography flame ionization detector,

2.2.2.3. Other extraction methods for OSCs in various matrices

Other sample preparation methods used for the extraction of OSCs prior to chromatographic analysis includes desorption, supercritical fluid extraction (SFE), reflux and the use of polymer thin film. Table 2.4 summarises all the other extraction and the most reported extraction method was the SFE. The latter uses supercritical fluid as a solvent for extraction and is suitable for both the solid and liquid matrices but mostly on the former. The commonly reported sample matrix on extraction of OSCs using SFE is coal. During SFE, supercritical fluid such as carbon dioxide (CO₂) is mostly used and the CO₂ is sometimes altered by co-solvent like ethanol or methanol [142]. However, toluene, 2-propanol and toluene-tetrahydrofuran are some of the organic solvents used in SFE. The latter was used by Gryglewicz and co-workers during the characterization of OSCs in coal extracts prior to GC-MS analysis. The combination of propanol and toluene as co-solvents had a good extraction yield of 39.9 wt.% when compared to other solvents. However, the use of CO₂ has advantages as it is non-corrosive, non- toxic and non-flammable [143]. Furthermore, SFE was used as an extraction method in coal and it was compared with ultrasonic assisted extraction (UAE) and accelerated solvent extraction (ASE). Different solvents for example, hexane and DCM were utilized for both UAE and ASE, while tetrahydrofuran (THF) and toluene combination were used for SFE. This resulted in better yield of 16% whilst ASE achieved 1.3% and UAE attained 1.4%. Although the ASE method achieved the lowest yield, the highest number (150) of OSCs were obtained by using this method [144]. However, the high pressure required for the removal of the supercritical fluid can be costly. Furthermore, CO₂ has its dissolving capacity limited by being a non-polar molecule [142].

The second popular extraction method was ligand exchange chromatography (LEC), which was assisted by PdCl₂ for the enrichment isolation of polycyclic aromatic sulphur compounds (PASHs) such as thiophenes (Th), benzothiophenes (BT) and dibenzothiophene (DBT) [143]. This method was as highly efficient and rapid for the separation of PASHs and PAH to avoid interferences when chromatographic analysis was done. The LEC with Pd (II) complex based on 2-amino-1-cyclopentene-1-dithiocaboxylate (ACDA) silica column was used for separation and it yielded excellent separation for the classes of compounds. Gas chromatography analysis of crude oil also showed that non-sulphur compounds were completely isolated [145]. Further

investigation was conducted on synthesized larger number of thiophene compounds using the Pd (II) complex. It was found that benzothiophenes are strongly retained when compared to thiophenes [146]. Other metals apart from Pd (II) were evaluated for the separation of OSCs. The ligand compounds such as CuSO₄, HgCl₂, AgNO₃, PdCl₂ (aq) and PdCl₂ were evaluated for the separation of OSCs. The study revealed that PdCl₂ (aq) had the best separation [147].

Polymer thin film is another pre-treatment method highlighted in **Table 2.4**. The polymer films are artificial materials formed from polymerization of monomers in the presence of a target analyte. These polymers are constant over a range of conditions including temperature and pressure [148]. A good reproducibility and a relative standard deviation value of 7.0% was achieved using this method for the selective analysis of thiophene compounds in water samples. During this study, the films were coupled to an HS-GC-SCD for the determination of semi-volatile compounds. Low LODs (0.24-0.82 μ g L⁻¹) with good linearity (0.99) and accuracy (89-98%) were obtained [58].

Sample	Detection Technique	Extraction method	Extraction time (min)	LOD (µgL ⁻ ¹)	Precision (%RSD)	Ref
Asphalt mixture	LC-MS	Reflux	ND	0.5 –	0.55 –	[150]
				346.5	16.01	
Coal	GC-MS	SFE	ND	-ND	-ND	[143]
Coal	GC-MS	SFE	ND	-ND	-ND	[151]
Coal	GC×GC-ToF-	SFE/ASE/UE	ND	-ND	-ND	[144]
	MS					
Crude oil	GC×GC-	Distillation	ND	-ND	-ND	[71]
	FID/SCD					
Gasoline/Heavy	Size exclusion	Ligand exchange	ND	ND	ND	[145]
distillates	chromatography					

 Table 2. 4 Indirect analysis of OSCs in different matrices using other sample

 pre-treatment followed by chromatographic techniques

Gas	oils	GC	Ligand exchange	ND	ND	ND	[147]
Petroleum			l igand exchange	ND	ND	ND	[146]
renoieum			Liganu exchange		ND		[140]

ND = not determined, LC-MS = liquid chromatography mass spectrometry, GC-MS = gas chromatography mass spectrometry, GC×GC-ToFMS = comprehensive two dimensional gas chromatography time-of-flight mass spectrometry,

2.3. Comparison of direct and indirect chromatographic analysis of OSCs

Direct and indirect chromatographic methods have been applied for analysis of OSCs in different samples using direct and indirect analysis. Figure 2.2A shows a number of literature publications that were reported for both direct and indirect chromatographic determination of OSCs from 2000 up to 2020 (two decades). Although the direct method eliminates many steps, thereby minimizing the contamination of the sample prior to analysis, it shows low number of publications as compared to indirect method. The observed trend might be due to the fact that, OSCs are commonly found in low concentration levels, thus sample pre-concentration is necessary. Furthermore, due to the complexity of the samples and the volatility of OSCs, sample pre-treatment is crucial, thus, a high number of publications for the indirect method are observed. During the first decade (2000-2010), at least eight publications were obtained on direct analysis of OSCs, followed by 11 publications on the second decade (2010-2020). A slight increase in publication numbers can be attributed to the availability of powerful analytical machines like GC×GC, which can give better separation of OSCs without the requirement of sample preparation steps. It is worthy to indicate that, only two publications described direct analysis of OSCs without sample dilution.

Different sample matrices that were analysed for chromatographic quantitative and qualitative determination of OSCs are illustrated in **Figure 2.2B**. From this figure it can be observed that, no publications were reported for the direct analysis of environmental (wastewater, sediments, and air) and food samples. The low concentrations of OSCs in these samples and the complexity nature of these samples requires extensive sample pre-treatment prior to OSCs chromatographic analysis. The least number of publications (2) was reported for biological sample matrices. It must be noted that these two publications were conducted at the beginning of the two decades, thus a lot of research still needs to be carried out for indirect and indirect determination of OSCs in these samples. Additionally, biological samples preferred direct analysis of OSCs. However, fossil fuel samples have the highest number of publications (16) for the direct approach and most of these samples had to undergo dilution prior to OSCs analysis. The reason for such a high number of publications might be that fossil fuels contain high concentration of OSCs, which can be easily detected without the need of extensive sample preparation.

However, **Figure 2.2B** also shows that, food matrices were mostly analysed for OSCs by using indirect chromatographic methods. The food samples include a wide range of scope such as cheese, beef, and drinks like beer. The main contributor to the highest number of publications in food was wine. This can be due to the aforementioned reasons in relation to the presence of OSCs causing undesirable flavours [161]. The category 'other' as reflecting in **Figure 2.2B**, includes samples such as Linchens and Lang baiju. Although these classes of matrices were reported to contain a wide range of OSCs, they favoured the indirect approach (7 publications) more than direct methods (2 publications).





Figure 2. 20 Number of publications for direct and indirect analysis of OSCs over two decades (A) and (B) various matrices

2.4. Comparison of extraction methods (LLE, SPE and other) for determination of OSCs

The indirect chromatographic analysis of OSCs in different matrices involved the use of various sample pre-treatment techniques. From the literature obtained, the sample pre-treatment techniques included solid phase extraction (SPE), liquid-liquid extraction (LLE), and other sample pre-treatment techniques. The 'other' sample pretreatment category includes techniques such as microwave assisted extractions (MAE), ultrasound assisted extraction UAE), stir bar sorptive extraction (SBSE), accelerated solvent extraction (ASE) and supercritical fluid extraction (SFE). Figure 2.3 shows that other sample pre-treatment methods have been less favoured for the extraction of OSCs, prior to chromatographic analysis. Furthermore, Figure 2.3 shows that SPE has the highest number of publications (22) from the past two decades. The highest number of publications for SPE might be due to its simplicity and the wide choice of adsorbents that can be used to improve extraction efficiencies of various OCSs. Additionally, SPE is rapid and minimizes the use of carcinogenic organic solvents that are commonly applied in LLE. The latter technique has three classes of micro techniques namely dispersive liquid-liquid microextraction (DLLME), headspace single drop microextraction (HS-SDME) and hollow fibre liquid-liquid microextraction (HF-LLME). These microextraction methods have immensely high number of publications for the LLE technique. This trend might be due to the use of microliter

solvents, which is low quantity when compared to the amount used during traditional LLE methods.



Figure 2. 3 Comparison of sample pre-treatment methods

2.5. The most analysed OSCs in various matrices

Sulfoxides, sulphides, thiophene, and benzothiophene were the reported OSCs in different samples (see **Figure 2.4**). However, sulfoxides were the least analysed compounds amongst the other three OSCs, with only 5 publications over the two decades. It has to be noted that, this analyte is not as toxic as the other analytes such as the polycyclic aromatic sulphur heterocycles (PASHs) and it is mostly reported in biological samples. Thiols were found in almost all the reported sample matrices; hence the highest number of publications (31) was observed for these sulphur compounds. Furthermore, fossil fuels were reported to be dominated by thiophenes. This can be attributed to the fact that, fossil fuel samples including crude oil are regarded as complex matrices, consist of various aromatic sulphur compounds, which are challenging to analysed, and remove. According to **Figure 2.4**, dibenzothiophene has been an analyte of focus for most researchers with more than 30 publications. This analyte and its derivatives have received much attention from researchers because they are one of the OSCs that contributes immensely to environmental pollution and mostly studied for development of desulphurization strategies.





2.6. Comparison of chromatographic techniques for analysis of OSCs

Figure 2.5 summarizes the used chromatographic techniques for the analysis of OSCs for the past two decades. No publications were reported in the year 2009. At the beginning of the first decades (first three years), there were no reports of GC×GC whilst reports on HPLC were published in 2000, 2002 and 2003 with only 1 publication per year. However, HPLC was also reported in 2014 with the highest number of publications (3) and again in 2016 with only 1 publication. This technique was only applied in sample matrices such as garlic, saliva, and water. Additionally, volatile OSCs were not suitable for HPLC, thus a low number of publications were published on this technique. The technique with low publications (4) overall is LC. All the publications reported on LC were done on the second decade, between 2015 and 2018. TLC has also been replaced by LC, because it does not differentiate between individual OSCs [157].

Alternatively, gas chromatographic techniques have been favoured and widely used for the determination of OSCs over the past decades. It has been noted that, GC was used in almost all the sample matrices, hence the high number of publications. Additionally, GC×GC was published more in the second decade with 10 publications, which is eight publications less compared to GC. Because of its increased selectivity towards OSCs and its enhanced power of separations using two columns, GC×GC has gained more popularity from researchers compared to GC [162]. Furthermore, due to the complex nature of the samples, GC cannot offer satisfactory separation especially in the presence of hydrocarbons [163], hence most studies are currently

focusing on the use of GC×GC.



Figure 2. 5 Chromatographic techniques used for OSCs analysis in the past two decades

EXPERIMENT AND METHODOLOGY

PREAMBLE

This chapter presents all the experimental procedures that were followed to generate the data reported and discussed in the next chapter. Standard operation procedures, the sample collection procedures, sample pre-treatment and sample analysis followed for both qualitative and quantitative analysis of OSCs are explained in detail. A gas chromatography coupled to a high resolute time-of-flight mass spectrometry (GC-TOFMS) used to generate data for the analysis of OSCs is also detailed in this chapter. Furthermore, different multivariate optimization procedures are discussed, including the one employed for this project. Lastly, physiochemical techniques (UV-VIS, TEM, SEM-EDX, FT-IR and P-XRD) used for the characterization of the proposed adsorbent are also explained in this chapter.

3.1 Standard operation procedure

All the laboratory experiments were conducted in glassware. The glassware was cleaned by soaking in acidic solution (3M HCl in ethanol) for a minimum of 3 hours, then rinsed three times in deionized water followed by drying in an oven overnight before use.

3.2 Sample collection

All crude oil samples were obtained from one of the South African petroleum company, while diesel, gasoline and kerosene samples were purchased from local filling stations.

3.3 Chemicals and standards

Analytical grade dichloromethane (DCM, ≥ 99.8) solvent, was purchased from Sigma-Aldrich (Johannesburg, South Africa

The following salts, iron (III) chloride hexahydrate (FeCl₃.6H₂O) ACS reagent, ferrous chloride tetrahydrate (FeCl₂.4H₂O), sodium citrate dihydrate \geq 99 % FG [HOC(COONa)(CH₂COONa)₂.2H₂O], tetrachloroauric acid (HAuCl₄.3H₂O) ~ 49% Au, and sodium hydroxide (NaOH) were also purchased from Sigma-Aldrich (Johannesburg, South Africa). Organosulphur standards, dibenzothiophene (98 %), 3-methylthiophene (98 %), thiophene (\geq 99%) and benzothiophene (95%) were also purchased from Sigma-Aldrich (Johannesburg, South Africa). Different solutions

including 28 w/v % ammonium solution, hydrochloric acid (98 %) was also purchased from Sigma-Aldrich (Johannesburg, South Africa). Diesel, kerosene and petroleum were purchased from a local filling station (South Africa). Nitric acid (65 %), hydrochloric acid (37 %) was supplied by Merck (South Africa). Ultrapure water which was used during the study with resistivity of 18.2 M Ω .m was obtained from a Milli-Q system. Glassware and GC vials were soaked in nitric acid (6 M) and then rinsed with ultra-pure water.

3.4 Preparation of standards

A 1000 ppm standard stock solution of dibenzothiophene (DBT) was prepared by weighing 10 mg (\pm 0.1 mg) of DBT. It was transferred into a clean, 10 mL volumetric flask that was half-filled with dichloromethane (DCM) and stored at -5 °C for later use. Working standards mixture of the target compounds were diluted with dichloromethane.





Figure 3. 1 Summary of the overall method

3.6 GC method and qualitative analysis

The prepared stock solution mixture was injected into the GC-TOFMS immediately after preparation. The different concentration (5, 10 and 15 ppm) prepared of the stock solution mixture was injected respectively in triplicate.. Crude oil, diesel, gasoline and kerosene samples were also screened for the OSCs by dissolving 100 μ L of each

sample in DCM as depicted in **Figure 3.2**. After dilution, each sample was injected in triplicates also.



Figure 3. 2 Qualitative analysis of OSCs in crude oil by dilution prior to GCToFMS analysis

3.7 Quantitative analysis

3.7.1 Preparation of Au-Fe₃O₄

Fe₃O₄ preparations

Chemical co-precipitation method was used for the synthesis of Fe₃O₄ nanoparticles. This method was selected because of its simplicity, low production costs, high yield and effectiveness compared to other methods such as microbial incubation which is slow and laborious [164]. There are two types of co-precipitation methods; the first involve the use of ferrous hydroxides and oxidizing agents. The co-precipitation method used was reported by Lancu (2020) et al . [166] and was followed with modifications. A 0.85 mL of conc. HCl (12 M) was diluted with 25 mL ultrapure H₂O. The prepared solution was used to dissolve 5.2 g of FeCl₃.6H₂O (ferric chloride hexahydrate) and 2 g of FeCl₂.4H₂O (ferrous chloride tetrahydrate). The Fe²⁺/Fe³⁺ ratio was important for the control of morphology and the size of the nanoparticles to be formed [167]. This mixture was conducted in an inert (N₂) environment. Then, 250

mL of NaOH (1.5 M) was added to the iron mixture. A black precipitate was formed and an external magnet then separated it at room temperature. The precipitate was washed four times using ultrapure H₂O. After washing, the back solid was again suspended in 250 mL of ultrapure H₂O mixed with 1 mL of NaOH (1.5 M) for 20 minutes. The synthesized Fe₃O₄-nanoparticles was stabilized by mixing it with 0.1 M (500 μ L) glucose [166].

Au-Fe₃O₄ preparations

The as-synthesized Fe₃O₄ NPs were covered with gold (Au) shell by first adding 20 mL of HAuCl₄.3H₂O (50 mM) into 105 mL of deionized water followed by boiling. Then, 5 mL (1mg/ mL) of the as-synthesized Fe₃O₄ was added into the boiling solution. Sodium citrate (5 mL, 80 mmol/L) was added slowly into the solution under constant stirring. The colour of the solution changed from brown to burgundy. The boiling continued for 5 minutes. The heat source was removed and stirring continued until the solution cooled to room temperature. Au-Fe₃O₄ was separated using a permanent magnet [168–170]

3.7.2 Magnetic solid phase extraction of organosulphur compounds

Blank extraction was performed using the solvent firstly. Then m-SPE procedure was done based on study by An (2018) et al. with modifications [171]. samples (10 mL) samples were placed in a beaker followed by 50 mg of Au-Fe₃O₄ nanoparticles. The mixture was then shaken for a certain period (10 min) to ensure complete preconcentration of the analyte. The adsorbent was then separated using an external magnet. The supernatant was decanted and an eluent (acetonitrile) of 100 μ L was introduced to desorb the analyte and mixed with the sample. Separation was done using an external magnet and the supernatant (1,5 ml) was collected and filtered using 0.22 μ m (13mm diameter) glass membrane/nylon filter purchased from Merck suppliers. After filtering, the supernatant containing a target analytes was analysed for OSCs using GC-TOFMS. **Figure 3.3**, summarizes the analysis of OSCs using m-SPE and GC-TFMS.



Figure 3. 3 Schematic representation of m-SPE for the quantitative analysis of OSCs in crude oil samples using GC-HR-ToF-MS

3.7.3 Multivariate optimisation *Screening*

The screening of the most significant variables was performed using the two level full factorial design (2⁴) with three replicates. Each factor was studied at two levels coded -1 for low level and +1 for high level. The factorial levels for the four variables, *viz*, extraction time, adsorbent mass, eluent volume and the pH were selected using a trial and error method. The data was assessed using Mini-Tab 18 software which produced Pareto charts that indicated significant factors at a certain confidence level based on the vertical line [172]. The significant parameters were further optimized using the response surface methodology outlined in the next section.

Response surface methodology

The second order response surface methodology was based on the central composite design (CCD). The significant parameters from the first order design were further optimized while the insignificant parameters were kept constant. The 3D surface plots from MiniTab 18 software were used to interpret analytical results [173]. On these plots, the relationship between the response and the parameters were formulated. Furthermore, the effect of each parameter on the extraction recoveries was observed [174].

3.7.4 Method validation

The m-SPE method was validated by assessing the percentage recovery, the precision in terms of the percentage relative standard deviation, linearity, limit of detection (LOD), limit of quantification (LOQ). A spike recovery strategy was employed for the evaluation of the accuracy at three concentration levels (6 μ g/g, 10 μ g/g ppm and 25 μ g/g) in triplicate to all the four samples (100 μ L). Calibration curves were constructed in order to assess the linearity by spiking ultra-pure water with a mixture of target analytes in the range concentration of 0,2 – 20 μ g/g for thiophene, 0,05 – 30 μ g/g for 3-methylthiophene, 2,6- 19 μ g/g for dibenzothiophene and 0,1 – 13 μ g/g for benzo[b]thiophene. Furthermore, the LOD and LOQ were calculated as:

$$LOD = 3SD/m$$
 and $LOQ = 10 SD/m$

Where SD is the standard deviation of the peak areas and m is the slope. These are reported on chapter 4.

3.8 Instrumentations

3.8.1 Gas chromatography coupled to a time-of-flight mass spectrometry (GC-ToFMS)

The samples were analysed using a LECO Pegasus High Throughput (HT) gas chromatographic time-of-flight mass spectrometry (GC-ToF-MS) (LECO corporation, St Joseph, Michigan, USA) system equipped with Agilent GC model 7890 (Agilent Technology, Santa Clara, California, USA). It was used for qualitative analysis of OSCs.. It was also fitted with jet-cooled modulator. A split/splitless injector set at 300 °C was also fitted. An HP-5m J&W capillary column with the dimension of 30 m × 0.25 mm × 0.25 µm film thickness with 5%- diphenyl/95% -dimethyl polysiloxane stationary phase supplied by Restek was used for separation. The instrument was controlled by and data acquisition was done using Leco Chroma TOF software . The oven conditions used for separation were as follows: The initial temperature was set at 50°C for a duration of 2 minutes until 140 °C. It was then ramped at 20 °C/ min until 200 °C for 2 minute. The ramping rate was increased to 30 °C/min and kept constant for 5 minutes until 260°C. The total analysis time was approximately 23 min.

Split/splitless type of front inlet was used and during the analysis, it was used in splitless mode with 1 μ L sample injection. The front inlet septum was operated on a flow of 3 mL min⁻¹ and the temperature of the injector was 300 °C. A carrier gas Helium

(99.999%, Afrox, Johannesburg, South Africa) was the carrier gas used under constant flow mode of 1 mL min⁻¹. The GC/ToF-MS interface temperature was set at 250 °C and mass spectra were obtained at 70 eV in full scan mode (scanning m/z ranging from 45 to 450). Data was analysed using Chroma ToF-HRT software.

The next section entails the synthesis of magnetic adsorbent for quantitative analysis of OSCs. Therefore, different characterization techniques including Fourier Transform Infrared spectroscopy (FTIR), Ultraviolet-visible spectrophotometry (UV-VIS), Scanning electron microscope (SEM), Transmission electron microscopy (TEM) and X-ray powder diffraction (P-XRD) were used to confirm the synthesized magnetic adsorbents.

3.8.2 Ultraviolet-visible spectrophotometry

The Genesys 180 UV-Visible spectrophotometer was used for confirmation of the nanoparticles of Fe₃O₄ and Au-Fe₃O₄. The dissolved solid samples (Fe₃O₄ and Au-Fe₃O₄) were first put in centrifuge tubes and diluted to make it less concentrated. Blank samples references were also prepared prior to analysis of real samples using distilled water. The samples were then run at a wavelength range of 200-1200 nm

3.8.3 Scanning electron microscope (SEM-EDX)

The morphology of the nanoparticles was confirmed used SEM-EDX. The scanning electron microscope (SEM) measurements were performed on a Tescan Vega 3 LMH at a voltage of 20 kV, with a secondary electron detector (SED) and energy dispersive X-ray spectroscopy (EDX). Prior to measurement, the samples were carbon-coated with the Agar Turbo Carbon coater to improve their conductivity

3.8.4 Fourier transform infrared spectroscopy (FT-IR)

The functional groups of the nano particles and its structural changes were investigated using the Fourier transform infrared (FTIR) spectra with a Bruker Tensor 27 FTIR spectrophotometer. The synthesized samples were mixed with potassium bromide (KBr) before being compressed into pellets. During the analysis, data was collected from 400 to 4000 cm⁻¹ region.

3.8.5 Powder X-ray diffraction (P-XRD)

The crystal structure of the nano particles was determined using the powder X-ray diffraction (P-XRD). The latter measurements were taken with a PANayltical X'Pert Pro powder diffractometer. The diffractometer was fitted with a 1D X'Celerator detector, upper levels of 6.67 and 12.78 keV, respectively, as well as programmable divergence slitting (10 mm radiation length). The measurements were taken with Cu K radiation in the 2 range from 5 to 90° at 40 kV and 40 mA operational conditions, the wavelength was 0.15405 nm. The diffractometer was set up with a sample spinner (Spinner PW3064) and a rotation time of 1 s to get the preferred orientation of crystallites. Predictions for the raw P-XRD patterns were made using High Score (Plus) software and ICDD PDF-4+ 2015.

3.8.6 Transmission electron microscopy (TEM)

The internal structure of the nano particles was studied using Transmission electron microscopy (TEM). It was carried out using a Jeol JEM-2100F Field Emission Electron Microscope at an acceleration voltage of 200 kV (JEOL Inc., Akishima, Japan) equipped with a LaB6 power source. The TEM samples was dispersed in methanol and a drop of the mixture was transferred onto a TEM grid (Cu-grid, 200 mesh) coated with a lacy carbon film.

RESULTS AND DISCUSSION

PREAMBLE

The experimental procedures outlined in the previous section (CHAPTER III) produced results that are detailed and discussed in this chapter. Furthermore, the characterization results of the synthesized gold coated magnetite used as an adsorbent are explained and discussed. Lastly, the results and data from a simple and efficient magnetic solid phase extraction (m-SPE) technique was developed under optimum conditions. This was followed by GC-ToFMS analysis for quantitative determination of organosulphur compounds (OSCs) in fuel oil samples including its figure of merits.

4.1 Qualitative analysis of OSCs using sample dilution

4.1.1 Total ion chromatogram

The identification of the OSCs was carried out via their nominal mass. The software was able to produce a total ion chromatogram (TIC) enabling visualization of the targeted OSCs.. The method allowed for the tentative determination of the OSCs including the thiophene isomer as observed on the peaks in **Figure 4.1**, which shows the TIC of targeted OSCs mixture of organic working standards. This mixture was used to develop the gas chromatographic method explained in **Chapter 3**. The retention times of these OSCs are also listed in Table 4.1. The chromatogram in Figure 4.1 shows good separation of benzothiophene and dibenzothiophene compounds. The study by Wilson and co-workers also produced similar peaks for polycyclic aromatic sulphur heterocycles although with use of normal-phase liquid chromatography (NPLC) fractionation prior to analysis [83]. It was anticipated that a co-eluation of polyaromatic hydrocarbons would be happen as explained on literature review. However, it appeared to be more prominent for compounds with high molecular mass or more substituted compounds to have a high retention time. The TIC in Figure 4.1 showed that dibenzothiophene was eluted late on the chromatogram because it is the most substituted OSCs, which clearly translates to high molecular mass because of the two benzene rings, attached, compared to the other compounds. The chromatogram also showed that thiophene was eluted very close to the solvent peak. This is due to the lower boiling point (85°C) of the compound that makes it to have a higher vapour pressure and shorter retention time since the compound spends much time in the gaseous phase [23]. Ruixiang and co-workers (2004) determined sulphur containing compounds in diesel oil by GC×GC-SCD and the retention times of the sulphur containing compounds also increased with an increase in their boiling points [15]. Furthermore, a study by Lopez Garcia (2002) also showed an increase on the retention times as the substituent on dibenzothiophene ring become bigger which consequently implied an increase on the molecular mass of the compound [24]. The selectivity of the HT-ToF-MS is comparable to that of sulphur chemiluminescence detector [25].





4.1.2 Mass spectrometry

The data from mass spectrometry is displayed in a form of a histogram as shown in **Figure 4.2**. This histogram gives an abundance of ions that reach the detection according to their mass charge ratio (m/z). All the peaks are normalized to the highest peak, i.e. m/z = 84, m/z = 97, m/z = 134 and m/z = 184 as indicated in the figure. The highest peaks are the molecular ion (primary ions) or the quantification peaks for all the target analytes.. This mass spectrum provided evidence of the presence of each

analyte. The molecular mass of thiophene for example is 84.14 Da. Figure 4.2 clearly shows the presence of this compound with a 96,9 % match when compared with the library spectra as indicated. The mass spectrum of dibenzothiophene showed isotopic charged molecular ion at m/z of 134, also providing evidence for the presence of dibenzothiophene. The mass spectra of the other compounds are also attached with Table 4.1 summarizing all the chromatogram and separation depicted in Figure 4.1. Many molecules have been analyzed by mass spectrometry before, and their spectra has been filed with the National Institute of Standards and Technology (NIST). The 2008 NIST library includes over 220,000 mass spectra. When the NIST library is available, it can be used to check for spectra that might match. Similar mass spectrometry was reported by Aloisi and co-workers where the similarity of compound identification was 906 for a substituted benzothiophene [18]. In this study, the similarity/match of benzothiophene was 931, which translated to above 93,1% when compared to the reported study and the other compounds showed a positive match or similarity with high percentage also (> 90 %) confirming the success of this method of analysis.



Caliper - sample "5PPM STD:1", 3:22.40 min:sec to 3:22.40 min:sec



Figure 4.2 Nominal mass spectrometry of individual OSCs from the mixture with library



Standards (5 ppm)	Retention time	Formula	Nominal	Mass
	(min)		mass	spectral
				similarity
Thiophene	3:22:40	C ₄ H ₄ S	84	974
3-methylthiophene	4:18:10	C_5H_6S	97	991
Benzo[b]thiophene	12:44:40	C_8H_6S	134	931
Dibenzothiophene	19:57:70	$C_{12}H_8S$	184	926

4.1.3 Qualitative analysis of real samples

The screening was conducted on real samples viz. crude oil, diesel, kerosene and gasoline. The analysis was conducted through diluting each sample and injecting it according to the chromatographic technique. Different OSCs compounds were tentatively identified in the samples. Figure 4.3-4.6 shows the chromatogram of the OSCs present in each sample. Compounds of interest were also present on the analysed samples (**Table 4.2**). The table also indicates that the compounds with low molecular mass are eluted at an early stage when compared to other compounds of high molecular masses. The tentative identification of these compounds on real samples was done with high similarity index. Table 4.2 indicates all the tentative identified compounds in the samples with some of their properties. A similar study of compound identification was conducted on jet oil using GC-AED/MS. The study reported an identification of 234 sulphides isomers [180].. This study identified 11 organosulphur compounds in kerosene sample, 10 OSCs in crude oil, 26 OSCs in diesel sample, 8 OSCs in gasoline with high similarities for some OSCs as indicated on **Table 4.2**. This clearly indicated that sample dilution could be used as an alternative method for analyte screening in different samples. Table 4.3 compares this study with other more studies conducted for screening or qualitative analysis of OSCs.



Figure 4. 3 TIC of OSCs contained in diesel sample



Figure 4. 4 TIC of OSCs contained in crude oil sample



Figure 4. 5 TIC of OSCs contained in gasoline



Figure 4. 6 TIC of OSCs compound contained in kerosene

Retention times (min)	OSCs	Molecular formula	Peak area	Molecular mass
	Diesel			
04:08,7	Thiophene, 3-methyl	C_5H_6S	433408	98,16
04:33,6	2-Thiophenecarboxylic acid, 4-nitrophenyl ester	C11H7NO4S	5747829	249,24
08:42,2	Acetic acid, mercapto-	$C_2H_4O_2S$	7796493	92,12
09:35,5	Thiazol-2-amine, N-(4-dimethylaminobenzyl)-	C12H15N3S	5400992	233,33
11:23,4	benzo[b]thiophene	C_8H_6S	7279128	134,2
19:05,5	Thieno[3,4-d]-1,3-dioxole, tetrahydro-2,2-dimethyl-, 5,5-dioxide	C7H12O4S	6391109	192,23
19:55,4	dibenzothiophene	$C_{12}H_8S$	663300	184,26
21:10,0	Sulfurous acid, octyl 2-propyl ester	C11H24O3S	8290728	236,37
22:45,7	Benz(1,4)oxathiino(2,3-c)pyridine	C11H7NOS	10097397	201,24
23:13,8	3-hydroxy-2-thiabutane	C ₃ H ₈ OS	68294015	92,15
24:12,2	2-Thiopheneacetic acid, isopropyl ester	C9H12O2S	1760520	184,25
24:26,4	3H-[1,3,4]Oxadiazole-2-thione, 5-(4-fluorophenyl)-3- (piperidin-1-yl)methyl-	C14H16FN3OS	49120968	293,36
25:53,4	2-Butanone, 3,3-dimethyl-1-thiocyanato-	C7H11NOS	10672795	157,23

Table 4. 2 Different OSCs contained in crude oil, kerosene, diesel and gasoline samples

26:25,0	3-Acetyl-2,5-dichlorothiophene	C ₆ H ₄ Cl ₂ OS	29360712	195,07
26:59,2	2(3H)-Thiazolethione, 4-methyl-	$C_4H_5NS_2$	49060908	131,22
27:06,2	1,3-Diphenyl-4H-1,2,4-triazoline-5-thione	$C_{14}H_{11}N_3S$	1877009	253,32
27:14,8	Benzothiazole, 2-(m-tolyl)-	$C_{14}H_{11}NS$	426584376	225,31
27:23,8	1,3-Diphenyl-4H-1,2,4-triazoline-5-thione	$C_{14}H_{11}N_3S$	2012999043	253,32
27:35,0	Ethylamine, N,N-dinonyl-2-phenylthio	C ₂₆ H ₄₇ NS	2198576738	405,72
27:56,3	1,3-Diphenyl-4H-1,2,4-triazoline-5-thione	$C_{14}H_{11}N_3S$	6521420	253,32
27:58,1	1,3-Diphenyl-4H-1,2,4-triazoline-5-thione	$C_{14}H_{11}N_3S$	1891188	253,32
28:01,3	10H-Phenothiazine, 2-(trifluoromethyl)-	C ₁₃ H ₈ F ₃ NS	443421	267,27
28:27,8	1,3-Diphenyl-4H-1,2,4-triazoline-5-thione	$C_{14}H_{11}N_3S$	178087613	253,32
28:35,2	1-(4-Methyl-dibenzothiophen-2-yl)-piperidine	C ₁₈ H ₁₉ NS	706473792	281,42
31:51,3	2,2-Dimethyl-propyl 2,2-dimethyl-propane-thiosulfinate	$C_{10}H_{22}OS_2$	669350617	222,41
32:35,8	Difluoro(methylamino)phosphine sulfide	CH ₄ F ₂ NPS	2015304	131,08
	Crude oil			
05:10,0	Thiophene, 2-(1,1-dimethylethyl)-	$C_8H_{12}S$	3009582	140,25
05:39,9	Thiophene, 3-methyl	C_5H_6S	293770482	98,16
08:37,7	Sulfurous acid, 2-ethylhexyl nonyl ester	$C_{17}H_{36}O_3S$	1313395738	320,53
09:52,3	Thiophene, 2-nitro-	$C_4H_3NO_2S$	3584020	129,14
09:56,9	Acetic acid, mercapto-	$C_2H_4O_2S$	1335228412	92,12

10:49,5	Benzenethiol, 4-(1,1-dimethylethyl)-	C ₁₀ H ₁₄ S	3598037	166,28
11:22,6	2-Thiophenecarboxylic acid, 5-tert-butyl-	$C_9H_{12}O_2S$	1336013527	184,26
12:33,6	benzo[b]thiophene	C_8H_6S	54414	134,2
13:15,4	4-(1-Methylpropylthio)pyrido[3,2-c]pyridazine	$C_{11}H_{13}N_3S$	24117800	219,31
13:22,4	Sulfurous acid, isobutyl pentyl ester	$C_9H_{20}O_3S$	388630153	208,32
13:37,6	dibenzothiophene	$C_{12}H_8S$	1908493197	184,26
	Gasoline			
03:48,7	Thiophene, 3-methyl	C₅H ₆ S	8339991	98,16
04:16,7	3-Picoline, 2-(tert-butylthio)-	$C_{10}H_{15}NS$	215040643	181,3
04:32,8	2-Thiophenecarboxylic acid, 4-nitrophenyl ester	C11H7NO4S	961027993	249,24
04:51,1	2-Thiophenecarboxylic acid, 4-nitrophenyl ester	C11H7NO4S	1192448932	249,24
09:11,4	2-p-Tolyl-N-(5-trifluoromethyl-[1,3,4]thiadiazol-2- yl)acetamide	C12H10F3N3OS	5759227	301,29
10:40,4	benzo[b]thiophene	C_8H_6S	83611	134,2
21:16,9	methanesulfonamide, N-[2-(octylamino)ethyl]-	$C_{11}H_{26}N_2O_2S$	27964937	250,41
22:40,6	dibenzothiophene	$C_{12}H_8S$	557880573	184,26
23:20,2	Tetramethyl diphosphane-oxide-sulfide	$C_4H_{12}OP_2S$	2489360204	170,15
24:05,6	Sulfurous acid, 2-ethylhexyl hexyl ester	$C_{14}H_{30}O_3S$	3135005990	278,45
	Kerosene			
05:10,0	Thiophene, 3-methyl	C_5H_6S	5445,8	98,16

08:37,7	Sulfurous acid, 2-ethylhexyl nonyl ester	C17H36O3S	17150017	320,53
09:52,3	Thiophene, 2-nitro-	$C_4H_3NO_2S$	660253668	129,14
09:56,9	Acetic acid, mercapto-	$C_2H_4O_2S$	2928689230	92,12
10:49,5	Benzenethiol, 4-(1,1-dimethylethyl)-	$C_{10}H_{14}S$	3526779253	166,28
11:22,6	2-Thiophenecarboxylic acid, 5-tert-butyl-	$C_9H_{12}O_2S$	5747829	184,26
	2-Ethyl-6-phenyl-1,3,4-thiadiazolo(3,2-a)(1,3,5)-		4302799781	
12:33,6	triazine-5,7-dione	$C_{12}H_{10}N_4O_2S$		274,3
13:15,4	benzo[b]thiophene	C_8H_6S	433408	134,2
13:22,4	Sulfurous acid, isobutyl pentyl ester	$C_9H_{20}O_3S$	526726563	208,32
	1-Butanesulfinamide, 1,1,2,2,3,3,4,4,4-nonafluoro-N-		2016139509	
13:37,6	methyl-	C ₅ H ₄ F ₉ NOS		297,14
19:24,1	dibenzothiophene	$C_{12}H_8S$	2720987822	184,26
4.1.4 Comparisons of studies

The study has showed satisfactory results when compared to other studies as indicated in **Table 4.4**. A total of 56 OSCs from the samples were screened with high degree of similarity above 50 % as indicated on **Table 4.3** without any pre-concentration technique employed. The number of compounds screened are equivalent to the study by Aloisi (2020) as indicated in **Table 4.4** which was done using a two dimensional gas chromatography.

Analytical	Sample matrix	Target OSCs	Total OSCs	Ref
instrument			screened	
GC-	Jet fuel	Thiols, sulphides and	-	[180]
AED/MS		thiophenes		
HPLC-	Crude oil	Thiophene	44	[181]
FTMS				
GC × GC-	Crude oil fractions	Thiols,	-	[71]
FID/SCD		benzothiophenes,		
		dibenzothiophene		
GC-Q-	Diesel fuels	Alkyldibenzothiophenes	19	[27]
TOFMS				
GC×GC-	Coal Tar	3-methylthiophene, 2,5-	60	[182]
HRToF-MS		dimethylthiophene, 2-		
		propylthiophene, 3-		
		butylthiophene,		
		benzo[b]thiophene, 2-		
		methylbenzo[b]thiophene,		
		dibenzothiophene (DBT),		
		4,6-		
		dimethyldibenzothiophene,		
GC-ToF-	Crude oil, Diesel,	Thiophene, 3-	19	This
MS	gasoline and	methylthiophene,		work
	kerosene	benzothiophene,		
		dibenzothiophene		

Table 4. 3 Comparison of different studies with current study

4.2 Characterization of nanoparticles

4.2.1 UV-VIS

Surface plasmon resonance (SPR) in gold nanoparticles was intense, resulting in an absorption peak in the visible portion of the electromagnetic spectrum as shown in **Figure 4.7**. The presence of superimposed peak represented magnetite and the red peak showed the presence of gold attached to magnetite.. This could be due to the reduced mean free path of the electrons for the small particles. The surface plasmon resonance is visible as a peak in **Figure 4.7** on the visible region, in the range between 520-530 nm. Elbialy et al.,[183]; Thao et al.,[184] and Keung et al., [170] have previously synthesized these nanoparticles and reported the presence of the peak at the same region reported in the current study.



Figure 4.7 UV-vis spectra of Fe₃O₄ and Au-Fe₃O₄ nanoparticles solution

4.2.2 Fourier transform infrared spectroscopy (FT-IR)

An IR spectra was used to confirm the identity of functional groups attached to the nanoparticles. The spectrums were compared to confirm the functionality. The two spectra in **Figure 4.8** showed similarities and that indicated that the functionality contained on both nanoparticles are similar in composition with major differences brought about the peaks at around 500 cm⁻¹. This peak in Fe₃O₄ is brought about the Fe-O stretching vibrational mode of Fe₃O₄. The peak is also present for the Au-Fe₃O₄ NPs but not sharp and intense because of the presence of Au. Figure 4.8 shows the peak between 1500 cm⁻¹ and 1750 cm⁻¹ can be assigned to the presence of C=O vibration. The presence of the C=O band can be attributed to citrate used during the synthesis, thus it can be associated with the -COOH group from the citrate shift. It also indicates the binding of the citrate to the Fe₃O₄ nanoparticles. Figure 4.8 also shows a sharp peak for both IR spectra at 3492 cm⁻¹. This peak can be assigned to the -OH groups brought about the presence of water on the surface of the nanoparticles. Stein and co-workers (2020) synthesized and characterized citrate stabilized gold-coated magnetite. During their study, characterization of the nano particles using the FT-IR showed a Fe-O between 550 cm⁻¹ and 650 cm⁻¹ [185] which is in line with what was obtained on current study. In another study, a deprotonated C=O peak was reported at 1710 cm⁻¹ [186] which is agreement with the current synthesis of the gold coated magnetite.





4.2.3 Scanning electron microscopy and energy dispersive x-ray (EDX) The prepared Fe₃O₄ and Au-Fe₃O₄ are characterised in Figure 4.9. The nanoparticles are well dispersed and uniform in size as observed by SEM, Figure 4.9A and C. These images show the spherical shapes of both the Fe₃O₄ and Au-Fe₃O₄ nano particles. Furthermore, EDX confirmed that the prepared nanoparticles contained the elements of Fe, C and O (Figure 4.9 B) for the synthesized magnetite. The detected signals of carbon arise from the SEM grid. The Au-Fe₃O₄ nano particles are confirmed with Figure 4.9 D with elemental distribution of Fe, O, C and Au at 55, 33.9, 8.8 and 1.8 wt. % respectively, which are elemental composition of Au-Fe₃O₄. The presence of Au element is not visible of the non-coated Fe₃O₄ as expected.



Figure 4.9 A) SEM for Fe₃O₄, B) EDX for Fe₃O₄, C) SEM for Au-Fe₃O₄ and d) EDX for Au-Fe₃O₄

4.2.4 Transmission electron microscopy

Transmission electron microscope was used to determine the shape and the size of the nanoparticles. The prepared Au-Fe₃O₄ nanoparticles mostly are spherical in shape, well dispersed and uniform in size as observed by TEM in **Figure 4.10**. The nanoparticles on the image look aggregated which can be due to the magnetic

properties it possesses. It also shows a shell in a form of gold that has surrounded the Fe₃O₄ nanoparticles. Due to the high electronic density of gold when compared to the magnetite, the shell structure of the particles are not visible on TEM, thus the employment of EDX on the previous section. The congruence of TEM with the SEM and EDX are confirmed as explained in the previous section. The Fe₃O₄ NPs were also spherical in shape with a diameter of about 10 nm. Previous studies of gold coated magnetite produced similar images [169].



Figure 4. 10 TEM for A) Fe₃O₄ B) Au-Fe₃O₄ core shell magnetic nanoparticles

4.2.5 Powder X-ray diffraction (P-XRD)

Powder X-ray diffraction patterns of Fe₃O₄ nanoparticles depict diffraction peaks (**Figure 4.11**) at 30.05, 35.33, 43.01, 53.39, 56.87, and 62.48u for which can be indexed to the (220), (311), (400), (422), (511), and (440) planes of Fe₃O₄ in a cubic phase. The diffraction peaks of Au-Fe₃O₄ nanoparticles are comparable to those of both Au and Fe₃O₄ nanoparticles. Their diffraction peaks can be assigned to Au (111), (200), (220), (311), and (222), and Fe₃O₄ (220), (311), (511) and (440) planes. The penetration of X-rays through the thin gold-coated layer to the central Fe₃O₄ core can reveal the diffraction peaks for Fe₃O₄. The diffraction patterns for both have been reported in the previous studies such as that conducted by Sedki and co-workers [187].



Figure 4. 11 P-XRD for Fe₃O₄ (black) and Au-Fe₃O₄ (red)

4.3 Quantitative analysis of OSCs using m-SPE extraction method4.3.1 Optimization of the extraction procedure

The screening process of most significant factors was conducted using a full factorial design with four independent variables. Mass of the sorbent, pH, eluent volume and time of extraction were chosen as the parameters to be varied. The minimum and maximum amount of each factor was chosen based on the previous studies. **Table 4.4** shows all the factors with the maximum and minimum amount for each factor.

Table 4. 4 Minimum and maximum amount used for different parameters fo	r
screening	

Variable	Minimum (-)	Maximum (+)
Mass of sorbent (mg)	10	100
Volume of Acetonitrile eluent (µL)	100	1000
Extraction time (min)	10	30
рН	4	9

The screening of significant parameters was done using atwo level full factorial design. The analytical response of each OSCs was given in a form of percentage recovery and the design matrix is displayed in **Figure 4.12**. The design of experiment (DOE) results produced Pareto charts form ANOVA that shows significant parameters on the preconcentration of OSCs using the magnetic solid phase extraction (m-SPE) procedure. The Pareto charts were constructed from four terms combination. The significant parameters are represented by the blue bars showed in **Figure 4.12** for each individual factor. The red-line indicates the 95% confidence level, thus from **Figure 4.12** it can be seen that extraction time is the only parameter that passed the 95% confidence level on thiophene, 3-methylthiophene and benzothiophene. Since the other parameters' bar length suggest that they are significant, further optimization is required thus the respond surface methodology is employed in the next section. Mass of the sorbent is another parameter which has a long bar length, thus in the next section, the two parameters i.e. extraction time and the mass of the sorbent will be further optimized to improve the recoveries of the OSCs extraction using the m-SPE.



Figure 4. 12 Full factorial design Pareto charts (n = 3) of A) thiophene, B) 3methylthiophene, c) dibenzothiophene and D) benzothiophene for the optimization of mass of sorbent, pH, eluent volume and extraction time at 95 % confidence level

Respond surface methodology

Central composite design (CCD) was used for robust mapping during the study in order to optimize further the parameters that are significant for the pre-concentration of the OSCs in crude oil samples. **Table 4.5** shows the central composite design matrix and its responses in terms of relative peak areas. The CCD deduced the interaction between the parameters under investigation and it helped in formulating the quadratic effects of the parameters, which are shown in equations 1-4. In this study, two experimental parameters were investigated at a two level full factorial design with 4 axial points and 5 centre points in cube. The experiments were conducted in triplicate for precision and accuracy. Each run produced 13 base runs with only base block. The central composite designs surface plots (**Figure 4.13 A-C**) were produced under optimum conditions in order to investigate the effect of mass of sorbent (40-120 mg) and the time of extraction (20-60 min). These conditions were informed by the results obtained from the previous section during the screening process. In **Figure 4.13** it can

be clearly seen that recoveries of above 75% for all the OSCs was obtained. For example, **Figure 4.13 c** shows that a recovery of more than 90% was obtained with a mass of 100 -200 mg and extraction time of around 60 minutes. The recoveries were satisfactory as they were above 75%. The conditions that yielded this satisfactory recovery were used in the next section for other sample pre-concentration using the magnetic solid phase extraction.

Thiophene = $-32,3 + 20,4 A + 0,943 B - 1,436 A^2 - 0,00462 B^2 - 0,0142 AB$ Equation 1 3 - methylthiophene = $18,8 + 9,2 A + 0,733 B - 0,780 A^2 - 0,00515 B^2 + 0,0156 AB$ Equation 2 Benzothiophene = $-26,6 + 24,1 A + 1.199 B - 1,393 A^2 - 0,00247 B^2 - 0,1369 AB$ Equation 3 Dibezothiophene = $-53,2 + 23,1A + 2,050 B - 1,44 A^2 - 0,00629 B^2 - 0,0927 AB$ Equation 4

	Experiment	Extraction time (min)	Mass of sorbent (mg)	Thiophene	3-methylthiophen	e Benzothiophene	Dibenzothiophene
•	1	40	110	80,3	79,3	87,9	90,1
	2	60	200	99,1	85,6	92,5	96,1
	3	40	110	68,5	73,6	77,9	81,3
	4	20	20	72,1	67,2	55,4	56,3
	5	11	110	77,2	75,4	89,3	91,3
	6	40	237	61,3	56,4	89,1	66,2
	7	68	110	50,2	70,8	70,1	55,5
	8	40	-17	0	0	0	0
	9	60	20	68,5	73,6	77,9	81,3
	10	40	110	70,2	67,4	69,3	70,9
	11	20	200	51,4	64,2	55,6	60
	12	40	110	71,9	77,5	81,9	90,5

 Table 4. 5 Respective percentage recoveries of central composite design matrix

13	40	110	77,9	71,9	83,9	76,5	
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Figure 4. 13 Central composite design optimization method for mass of sorbent and extraction time

4.3.2 Validation of magnetic solid phase extraction method

The proposed m-SPE method was validated by using five concentration levels of spiked and unspiked crude oil samples with three replications for each concentration level. The spike concentration levels for the sample were 6, 10, 17, 22 and 25 ppm. The concentration of the samples was plotted with peak areas to provide method calibration. The calibration was used to deduce figures of merit such as the linearity, precision, limit of detection, limit of quantification and the linear equations in order to validate the m-SPE on the OSCs analysis by GC-ToF-MS. The validation data is summarized in **Table 4.6.** The correlation factor for the proposed procedure ranged from 0,9816-0,9961 for all the investigated OSCs. Furthermore, the limit of detection and the limit of quantification ranged from 0,02-0,199 and 0,602-0,126 - 0,602 ug/g respectively. The precision of the method was also validated through the relative

standard deviation as it was in the range of 0,9-2,3 %. Since the RSD is less than 4%, this indicates that the method relatively has excellent reproducibility.

Compound	LODs	LOQs	linear correlation	Recovery	Linear RSD %
	(µg/g)	(µg/g)	coefficients(R ²)	(%)	equation
Thiophene	0,02	0,131	0,9816	76	y = 2E+06x + 2E+06 1,8
2 Mathulthianhana	0.0265	0.0906	0.002	02	
3-methylthiophene	0,0265	0,0806	0,993	93	y = 4E + 07X + 8E + 07 1,1
Benzothiophene	0,199	0,602	0,9961	88	y = 1E+08x + 7E+08 0,8
Dibenzothiophene	0,0416	0,126	0,9829	95	y = 1E+08x + 1E+09 2,3
					-

Table 4. 6 Figures of analytical merits for OSCs analysis in fuel oil samples at 25 μ g/g

LOD, limit of detection for = 3SD/m; LOQ limit of quantification = 10SD/m for S/N =10; RSD, relative standard deviation (n = 3)

4.3.3 Comparison with reported literature

Several reports have been compiled on different extraction and pre-concentration methods for the OSCs in different samples. **Table 4.7** gives the summary of the analytical figure of merits of the compiled reports with those of the current study. From the complied tabled, the current study has linearity of 0,9816-0,9961. Furthermore, it has a relatively good precision (0,8-2,3) when compared to other study such as study by , Yang et al [38]. The developed method can be used as an alternative with high precision for the analysis of OSCs. The lowest concentration of OSCs that can be detected was found to be between 0.08-0.602 μ g/g.. Although the limit of detection is higher, the methods that yield relatively low limit of detection requires high extraction time, use of commercially available fibres and low selectivity of the analyte [186]. Thus, the extraction time of the current study is lower which makes a rapid and low-cost method since the adsorbent can be easily synthesized.

Samples	OSCs	Method	Chromatography	LODs	%	Correlation	RSD	Ref
				(ppm)	Recovery	R ²	%	
Crude oil and	Benzothiophene, dibenzothiophene	LPME	GC-SCD	0,001 -	77 -91	0,98 – 0,99	4,5- 13,8	[36]
diesel	and derivatives			0,111				
Crude oil	Benzothiophene, dibenzothiophen, and benzo[b]naphtho[1,2- d]thiophene and their derivatives	µ-SPE	GC-MS		81.5 -92.1	0.9894 to 0.9994	3.23 – 7,52%	[38]
Coal tar	Benzo[b]thiophene, Thiophene and its derivatives	Dilution	GC × GC-ToF- MS	8,0 – 30,0	-	0,97414 – 0,9882	1 – 4	[182]
Crude oil	Thiophene, dibenzothiophen, 3-	m-SPE	GC-HR-ToF-MS	0,02 - 0,199	76 - 95	0,9816 – 0,9961	0,8 – 2,3	This work

Table 4. 7 Comparison of analytical merits for developed method with reported literature

methylthiophene and benzathiophene

4.3.4 Application of m-SPE in real fuel oil samples

The developed extraction method was then applied on diesel, gasoline and kerosene samples to detect the amount of OSCs present including the target analytes. This was conducted using the previously deduced optimum conditions applied on crude oil for validation. The amount of these OSCs was detected in some samples as depicted in **Table 4.8**. Furthermore, it can clearly be seen that the semi-quantification of kerosene sample contains the highest concentration of the OSCs ranging from 1,01-3,6 ppm. Crude oil sample was the second highest with semi- quantified concentration of OSCs ranging from 0,13-4,09 ppm. On the other hand, the kerosene sample did not show any presence of a lot of OSCs compounds as compared to the other samples. During the analysis of the samples, other OSCs compounds, which were not a target during the study, were visible.

Organosulphur compounds	Crude oil	Diesel	Gasoline	Kerosene
	(ppm)	(ppm)	(ppm)	(ppm)
Acetic acid, mercapto-	$0,3 \pm 0,005$	0,02 ± 0,001	1,06 ± 0,007	ND
Disulfide, diphenyl	ND	0,1 ± 0,02	3,06 ± 0,02	ND
Benzo[b]naphtho[2,3-d]thiophene, 9,10-dihydro-6,7- dimethyl-	1,06 ± 0,004	0,012 ± 0,1	3,05±0,04	1,01 ± 0,05
Disulfide, bis(4-methylphenyl)	ND	0,41 ± 0,06	ND	5,02 ± 0,0001
Benzenemethanethiol, TMS derivative	0,13 ± 0,08	ND	$0,46 \pm 0,003$	$3,6 \pm 0,2$
2-Thiophenecarboxylic acid, 5-tert-butyl-	1,02 ± 0,05	3,06 ± 0,03	2,06 ± 0,01	1,9 ± 0,003
Acetic acid, mercapto-	1,93 ± 0,03	ND	ND	ND
Benzo[b]thiophene, 3-phenyl-	$0,35 \pm 0,02$	2,15 ± 0,009	ND	2,45 ± 0,004
Thiophene, 2-iodo-	2,05 ± 0,07	ND	$1,04 \pm 0,005$	ND
5-Bromo-2-thiophenecarboxaldehyde	4,09 ± 0,1	2,01 ± 0,003	2, 65 ± 0,03	1,43 ± 0,4
Thioxanthene	ND	$0,9 \pm 0,006$	1,12 ± 0,05	ND

Table 4. 8 Semi-quantified Concentration of OSCs in crude oil, diesel, gasolineand kerosene samples using m-SPE followed by GC-ToF-MS

Experimental conditions: volume of eluent, 100 μ L; mass of sorbent, 150 mg; time of extraction, 50 min; and pH of sample, 6,7.

CONCLUSION AND RECOMMENDATION

5.1 Conclusion

. Sample dilution and an intensive magnetic solid phase extraction (m-SPE) were the methods developed. The sample dilution method was used for screening of the different OSCs contained in crude oil, diesel, gasoline and kerosene samples thus qualitative analysis.

During the qualitative analysis of the OSCs in the fuel oils, dichloromethane (DCM) was used as a solvent for the sample dilution prior to chromatographic analysis. The good properties of the solvent such as the good solubility of the solvent made it easy for the sample to be well dissolved. Moreover, DCM has a low boiling point and low molecular mass, thus peak interference between the analyte of interest and the solvent peak was eliminated. A total of 56 OSCs were obtained through screening of the samples. These compounds include thiols, disulphides, thiophenes and polycyclic aromatic sulphur heterocycles, just to name the few.

The quantitative analysis of OSCs in crude oil, diesel, gasoline, and kerosene was performed using the m-PSE procedure for preconcentration method prior to chromatographic analysis. During the m-SPE, a magnetic sorbent in a form of gold coated magnetite nano particles was used. The synthesized nano particles were successfully characterized to confirm the formation of the Au-Fe₃O₄ nano particles. UV-Vis peak at around 520 – 5230 nm which is a typical characteristic of Au or Fe3O4 was observed. The SEM and TEM showed that the particles have a spherical shape with particle size of 100 nm, while XRD showed that they have magnetite phase. FTIR showed Fe-O peak at 500 cm⁻¹ which proved that Fe₃O₄ nanoparticles were successfully synthesized Moreover, factors affecting m-SPE were evaluated using full factorial design. These were the volume of the eluent (100-1000 µl), the pH of the sample (4-9), the extraction time (10-30 min) and the mass of the adsorbent (10-100 mg). Through Pareto charts produced from the full factorial design, it was found that extraction time and mass of adsorbent were the most significant parameters. These significant factors were further optimized using the central composite design (CCD) as the response surface methodology to find the optimum conditions. The optimum conditions were found to be 150 mg for the mass of sorbent and 50 min for the time of

extraction. The other conditions that were kept constant were eluent volume which was 100 μ L of acetonitrile and the pH of the sample at 6.7.

Coupling the GC-ToF-MS with the developed sample pre-treatment method yielded an improved correlation factor of 0,9816-0,9961, better sensitivity of 0,8-2,3 % and acceptable recoveries of 76-95 %. The limit of quantification and the limit of detection were found to be in the range of 0,126-0,602 ppm and 0,02-0,0416 ppm respectively. The developed methods can be used as alternative methods for determination of OSCs in oily matrices. Upon analysing, the concentration levels of OSCs in crude oil, gasoline, diesel, and kerosene were found to be in ranged of 0,3-4,09; 1,06-1,65; 0,02-3,06 and 1,01-5,02 ppm, respectively. The concentrations of OSCs in the investigated fuel samples were in acceptable level according to the South African Petroleum regulations. The dilution method was rapid, cost-effective, and environmentally friendly, while m-SPE was simple and selective toward OSCs thus recommended for qualitative and semi-quantitative analysis of OSCs.

5.2 Recommendations for future studies

. In future, a comparison between this study and a study using a two-dimensional gas chromatography can be conducted to compare these studies since this instrument provides an extra dimension due to the presence of the dual columns. The number of targeted analytes can be increased to include more of the polycyclic aromatic sulphur compounds, since they contain different substituents, which can cause interferences during analysis. Furthermore, other parameters can be investigated instead of the four that this study focused on, this includes factors as such as the spiking concentration, different eluent etc. Other studies can also incorporate the use of a bio-adsorbent for the m-SPE because they are also inexpensive and ecologically friendly. These bio-adsorbents can be further used to remove the OSCs contained in different oily matrices.

- [1] F. Khalilian and M. Rezaee, "Extraction and determination of organosulfur compounds in water samples by using homogeneous liquid-liquid microextraction via flotation assistance-gas chromatographyflame ionization detection," *Bull. Chem. Soc. Ethiop.*, vol. 28, no. 2, pp. 195–204, 2014, doi: 10.4314/bcse.v28i2.4.
- K. A. Kvenvolden and C. K. Cooper, "Natural seepage of crude oil into the marine environment," *Geo-Marine Lett.*, vol. 23, no. 3–4, pp. 140–146, 2003, doi: 10.1007/s00367-003-0135-0.
- [3] R. C. Selley and S. A. Sonnenberg, "The Physical and Chemical Properties of Petroleum," *Elem. Pet. Geol.*, pp. 13–39, 2015, doi: 10.1016/b978-0-12-386031-6.00002-3.
- [4] M. Denchak, "Fossil Fuels: The Dirty Facts | NRDC," Natural Resources Defense Council. pp. 1–26, 2018, [Online]. Available: https://www.nrdc.org/stories/fossil-fuels-dirty-facts.
- [5] A. T. Jarullah, I. M. Mujtaba, and A. S. Wood, "Improvement of the middle distillate yields during crude oil hydrotreatment in a trickle-bed reactor," *Energy and Fuels*, vol. 25, no. 2, pp. 773–781, 2011, doi: 10.1021/ef101327d.
- [6] E. Summary, "The nature of petroleum reservoirs," *Dev. Pet. Sci.*, vol. 16, no.
 C, pp. 155–178, 1983, doi: 10.1016/S0376-7361(08)70092-X.
- [7] G. Yasin *et al.*, "Quality and chemistry of crude oils," *J. Pet. Technol. Altern.*, vol. 4, no. March, pp. 53–63, 2013, doi: 10.5897/JPTAF12.025.
- [8] F. Trejo and J. Ancheyta, "Characterization of asphaltene fractions from hydrotreated maya crude oil," *Ind. Eng. Chem. Res.*, vol. 46, no. 23, pp. 7571– 7579, 2007, doi: 10.1021/ie0700213.
- [9] Hot Lava Mobile Sample Apps, "Characteristics of Crude Oil," vol. 6, p. 2006,
 2006, [Online]. Available: http://hdwap.com/demo1/courseview/1.html?course=13248&pageId=1&f.
- [10] D. Stratiev, R. Dinkov, N. Nikolaev, and K. Stanulov, "Evaluation of impact of

crude oil quality on refinery profit," *Erdoel Erdgas Kohle*, vol. 126, no. 1, pp. 17–21, 2010.

- S. U. Siddiqui and K. Ahmed, "Methods for Desulfurization of Crude Oil-a Review," *Sci.Int.(Lahore)*, vol. 28, no. 2, pp. 1169–1173, 2016.
- [12] G. Coast and M. Europe, "Oil Refineries," *Oil Refineries*, no. April 2014, pp. 1– 11, 2015.
- [13] R. E. Correa Pabón and C. R. de Souza Filho, "Crude oil spectral signatures and empirical models to derive API gravity," *Fuel*, vol. 237, no. May 2018, pp. 1119–1131, 2019, doi: 10.1016/j.fuel.2018.09.098.
- Y. Han, Y. Zhang, C. Xu, and C. S. Hsu, "Molecular characterization of sulfurcontaining compounds in petroleum," *Fuel.* 2018, doi: 10.1016/j.fuel.2018.02.110.
- [15] J.G. Speight, handbook of petroleum refining, CRC Press, NY, 2017
- [16] J. G. Speight, "Petroleum: Chemistry, Refining, Fuels and Petrochemicals -Product Treating," *Encycl. Life Support Syst.*, vol. 1, pp. 8–13, 2003, doi: 10.1038/198049a0.
- [17] M. Soleimani, A. Bassi, and A. Margaritis, "Biodesulfurization of refractory organic sulfur compounds in fossil fuels," *Biotechnology Advances*. 2007, doi: 10.1016/j.biotechadv.2007.07.003.
- [18] R. Mishra, I. Tomar, S. Singhal, and K. K. Jha, "Synthesis, properties and biological activity of thiophene: A review," *Der Pharma Chem.*, vol. 3, no. 4, pp. 38–54, 2011.
- [19] J. H. Kareem, "Sulfur extraction from oil using ionic liquids by," no. June, p. 179, 2017.
- [20] S. C. Moldoveanu, "Pyrolysis of Thiols and Sulfides," *Pyrolysis Org. Mol.*, pp. 321–325, 2019, doi: 10.1016/b978-0-444-64000-0.00007-x.
- [21] R. Javadli and A. de Klerk, "Desulfurization of heavy oil," *Appl. Petrochemical Res.*, vol. 1, no. 1–4, pp. 3–19, 2012, doi: 10.1007/s13203-012-0006-6.
- [22] Nour Shafik El-Gendy, J.G Speight, Handbook of refinery desulfirization, CRC

Press, 2015, doi: 10.1201/b19102-4.

- [23] S. D. Emsbo-mattingly and E. Litman, Second handbook oil spill environmental forensics, 2nd, Academic press, 2016, p 255 - 312
- [24] H. C. Menezes, B. P. Paulo, M. J. N. Paiva, and Z. L. Cardeal, "A Simple and Quick Method for the Determination of Pesticides in Environmental Water by HF-LPME-GC/MS," *J. Anal. Methods Chem.*, vol. 2016, 2016, doi: 10.1155/2016/7058709.
- [25] F. Cappelli Fontanive, É. A. Souza-Silva, J. Macedo da Silva, E. Bastos Caramão, and C. Alcaraz Zini, "Characterization of sulfur and nitrogen compounds in Brazilian petroleum derivatives using ionic liquid capillary columns in comprehensive two-dimensional gas chromatography with time-offlight mass spectrometric detection," *J. Chromatogr. A*, vol. 1461, pp. 131–143, 2016, doi: 10.1016/j.chroma.2016.07.025.
- [26] R. A. Gimeno, A. F. M. Altelaar, R. M. Marcé, and F. Borrull, "Determination of polycyclic aromatic hydrocarbons and polycylic aromatic sulfur heterocycles by high-performance liquid chromatography with fluorescence and atmospheric pressure chemical ionization mass spectrometry detection in seawater and sediment samp," *J. Chromatogr. A*, vol. 958, no. 1–2, pp. 141–148, 2002, doi: 10.1016/S0021-9673(02)00386-2.
- [27] X. Zhu, M. Liu, Z. Liu, Y. Li, and S. Tian, "Direct and Rapid Quantitative Analysis of Alkyldibenzothiophenes in Deeply Hydrodesulfurized Diesel Fuel by Gas Chromatography Quadrupole Time-of-Flight Mass Spectrometry," *Energy and Fuels*, vol. 31, no. 9, pp. 9125–9131, 2017, doi: 10.1021/acs.energyfuels.7b01289.
- [28] G. Vanini *et al.*, "Analytical advanced techniques in the molecular-level characterization of Brazilian crude oils," *Microchem. J.*, vol. 137, pp. 111–118, 2018, doi: 10.1016/j.microc.2017.10.003.
- [29] J. Beens and R. Tijssen, "The Characterization and Quantitation of Sulfur-Containing Compounds in (Heavy) Middle Distillates by LC-GC-FID-SCD," *HRC J. High Resolut. Chromatogr.*, vol. 20, no. 3, pp. 131–137, 1997, doi: 10.1002/jhrc.1240200303.

- [30] F. A. Franchina, M. E. Machado, P. Q. Tranchida, C. A. Zini, E. B. Caramão, and L. Mondello, "Determination of aromatic sulphur compounds in heavy gas oil by using (low-)flow modulated comprehensive two-dimensional gas chromatography-triple quadrupole mass spectrometry," *J. Chromatogr. A*, vol. 1387, pp. 86–94, 2015, doi: 10.1016/j.chroma.2015.01.082.
- [31] G. D. da Silveira *et al.*, "A liquid chromatography–atmospheric pressure photoionization tandem mass spectrometric method for the determination of organosulfur compounds in petroleum asphalt cements," *J. Chromatogr. A*, vol. 1457, pp. 29–40, 2016, doi: 10.1016/j.chroma.2016.06.003.
- [32] W. B. Wilson, H. V. Hayes, A. D. Campiglia, and S. A. Wise, "Qualitative characterization of three combustion-related standard reference materials for polycyclic aromatic sulfur heterocycles and their alkyl-substituted derivatives via normal-phase liquid chromatography and gas chromatography/mass spectrometry," *Anal. Bioanal. Chem.*, vol. 410, no. 17, pp. 4177–4188, 2018, doi: 10.1007/s00216-018-1065-z.
- [33] S. Chiaberge, T. Fiorani, and P. Cesti, "Methyldibenzothiophene isomer ratio in crude oils: Gas chromatography tandem mass spectrometry analysis," *Fuel Process. Technol.*, vol. 92, no. 11, pp. 2196–2201, 2011, doi: 10.1016/j.fuproc.2011.07.011.
- [34] M. Mei, K. K. (Adry. Bissada, T. B. Malloy, L. M. Darnell, and E. B. Szymcyk, "Improved method for simultaneous determination of saturated and aromatic biomarkers, organosulfur compounds and diamondoids in crude oils by GC– MS/MS," Org. Geochem., vol. 116, pp. 35–50, 2018, doi: 10.1016/j.orggeochem.2017.09.010.
- [35] D. M. Jones, J. S. Watson, W. Meredith, M. Chen, and B. Bennett,
 "Determination of naphthenic acids in crude oils using nonaqueous ion exchange solid-phase," *Anal. Chem.*, vol. 73, no. 3, pp. 703–707, 2001, doi: 10.1021/ac000621a.
- [36] I. Al-Zahrani, C. Basheer, and T. Htun, "Application of liquid-phase microextraction for the determination of sulfur compounds in crude oil and diesel," *J. Chromatogr. A*, vol. 1330, pp. 97–102, 2014, doi:

10.1016/j.chroma.2014.01.015.

- [37] G. D. da Silveira *et al.*, "A cleanup method using solid phase extraction for the determination of organosulfur compounds in petroleum asphalt cements," *Fuel*, vol. 202, pp. 206–215, 2017, doi: 10.1016/j.fuel.2017.04.020.
- [38] B. Yang, W. Hou, K. Zhang, and X. Wang, "Application of solid-phase microextraction to the determination of polycyclic aromatic sulfur heterocycles in Bohai Sea crude oils," *J. Sep. Sci.*, vol. 36, no. 16, pp. 2646–2655, 2013, doi: 10.1002/jssc.201300184.
- [39] S. N. Nabavi, S. M. Sajjadi, and Z. Lotfi, "Novel magnetic nanoparticles as adsorbent in ultrasound-assisted micro-solid-phase extraction for rapid preconcentration of some trace heavy metal ions in environmental water samples: desirability function," *Chem. Pap.*, vol. 74, no. 4, pp. 1143–1159, 2019, doi: 10.1007/s11696-019-00954-z.
- [40] D. Huang, C. Deng, and X. Zhang, "Functionalized magnetic nanomaterials as solid-phase extraction adsorbents for organic pollutants in environmental analysis," *Anal. Methods*, vol. 6, no. 18, pp. 7130–7141, 2014, doi: 10.1039/c4ay01100g.
- [41] H. Lu, Q. Shi, J. Lu, G. Sheng, P. Peng, and C. S. Hsu, "Petroleum sulfur biomarkers analyzed by comprehensive two-dimensional gas chromatography sulfur-specific detection and mass spectrometry," *Energy and Fuels*, vol. 27, no. 12, pp. 7245–7251, 2013, doi: 10.1021/ef401239u.
- [42] H. Al-rabiah, L. Al-ostad, T. Al-shamary, A. Al-hendi, H. Al-henayyan, and R. Khazemi, "Sulfur speciation of Kuwaiti crude oils fractions using twodimensional gas chromatography," *Arab. J. Chem.*, 2019, doi: 10.1016/j.arabjc.2019.11.007.
- [43] S. S. Mgiba, V. Mhuka, and N. Mketo, "Trends in the Direct and Indirect Chromatographic Determination of Organosulfur Compounds in Various Matrices Trends in the Direct and Indirect Chromatographic Determination of Organosulfur Compounds in Various Matrices," *Sep. Purif. Rev.*, vol. 00, no. 00, pp. 1–13, 2021, doi: 10.1080/15422119.2020.1866011.

- [44] F. Bianchini and H. Vainio, "Allium vegetables and organosulfur compounds: Do they help prevent cancer?," *Environ. Health Perspect.*, vol. 109, no. 9, pp. 893–902, 2001, doi: 10.1289/ehp.01109893.
- [45] M. Mestres, O. Busto, and J. Guasch, "Analysis of organic sulfur compounds in wine aroma," *J. Chromatogr. A*, vol. 881, no. 1–2, pp. 569–581, 2000, doi: 10.1016/S0021-9673(00)00220-X.
- [46] R. S. Amais, G. L. Donati, and J. A. Nóbrega, "Sulfur determination in fuels by ICP-OES and ICP-MS to meet increasingly stricter legislation requirements," *Spectrosc. (Santa Monica)*, vol. 29, no. 3, 2014.
- [47] J. S. Sinninghe Damstě, C. M. White, J. B. Green, and J. W. De Leeuw,
 "Organosulfur compounds in sulfur-rich raša coal," *Energy and Fuels*, vol. 13, no. 3, pp. 728–738, 1999, doi: 10.1021/ef980236c.
- [48] C. Cerella, M. Kelkel, E. Viry, M. Dicato, C. Jacob, and M. Diederich, "Naturally Occurring Organic Sulfur Compounds: An Example of a Multitasking Class of Phytochemicals in Anti-Cancer Research," *Phytochem. - Bioactivities Impact Heal.*, no. December 2011, pp. 2–42, 2011, doi: 10.5772/26003.
- [49] R. T. Ruhee, L. A. Roberts, S. Ma, and K. Suzuki, "Organosulfur Compounds: A Review of Their Anti-inflammatory Effects in Human Health," *Front. Nutr.*, vol. 7, no. June, pp. 1–11, 2020, doi: 10.3389/fnut.2020.00064.
- [50] N. Muizzuddin and R. Benjamin, "Beneficial Effects of a Sulfur-Containing Supplement on Hair and Nail Condition," *Natu*, vol. 11, no. 11, pp. 1–8, 2019.
- [51] S. Fukushima, N. Takada, T. Hori, and H. Wanibuchi, "Cancer prevention by organosulfur compounds from garlic and onion," *J. Cell. Biochem.*, vol. 67, no. SUPPL. 27, pp. 100–105, 1997, doi: 10.1002/(SICI)1097-4644(1997)27+<100::AID-JCB16>3.0.CO;2-R.
- [52] H. R. Cadorim, É. R. Pereira, E. Carasek, B. Welz, and J. B. De Andrade,
 "Determination of sulfur in crude oil using high-resolution continuum source molecular absorption spectrometry of the SnS molecule in a graphite furnace," *Talanta*, vol. 146, pp. 203–208, 2016, doi: 10.1016/j.talanta.2015.07.088.
- [53] S. H. Omar and N. A. Al-Wabel, "Organosulfur compounds and possible

mechanism of garlic in cancer," *Saudi Pharm. J.*, vol. 18, no. 1, pp. 51–58, 2010, doi: 10.1016/j.jsps.2009.12.007.

- [54] A. Hall, A. Troupin, B. Londono-Renteria, and T. M. Colpitts, "Garlic organosulfur compounds reduce inflammation and oxidative stress during dengue virus infection," *Viruses*, vol. 9, no. 7, 2017, doi: 10.3390/v9070159.
- [55] Ž. Fredotović, B. Soldo, M. Šprung, Z. Marijanović, I. Jerković, and J. Puizina, "Comparison of organosulfur and amino acid composition between triploid onion Allium cornutum clementi ex visiani, 1842, and common onion Allium cepa L., and evidences for antiproliferative activity of their extracts," *Plants*, vol. 9, no. 1, 2020, doi: 10.3390/plants9010098.
- [56] N. Ochiai, K. Sasamoto, and K. MacNamara, "Characterization of sulfur compounds in whisky by full evaporation dynamic headspace and selectable one-dimensional/two-dimensional retention time locked gas chromatographymass spectrometry with simultaneous element-specific detection," *J. Chromatogr. A*, vol. 1270, pp. 296–304, 2012, doi: 10.1016/j.chroma.2012.11.002.
- [57] O. Busto, J. Guasch, M. Mestres, and P. Mart, "Simultaneous analysis of thiols, sulphides and disulphides in wine aroma by headspace solid-phase microextraction–gas chromatography ´," *J. Chromatogr. A*, vol. 849, pp. 293– 297, 1999.
- [58] H. Y. Hijazi and C. S. Bottaro, "Selective determination of semi-volatile thiophene compounds in water by molecularly imprinted polymer thin films with direct headspace gas chromatography sulfur chemiluminescence detection," *Analyst*, vol. 143, no. 5, pp. 1117–1123, 2018, doi: 10.1039/c7an01686g.
- [59] R. S. Amais, C. D. B. Amaral, L. L. Fialho, D. Schiavo, and J. A. Nóbrega,
 "Determination of P, S and Si in biodiesel, diesel and lubricating oil using ICP-MS/MS," *Anal. Methods*, vol. 6, no. 13, pp. 4516–4520, 2014, doi: 10.1039/c4ay00279b.
- [60] Y. Han, Y. Zhang, C. Xu, and C. S. Hsu, "Molecular characterization of sulfurcontaining compounds in petroleum," *Fuel*, vol. 221, no. February, pp. 144– 158, 2018, doi: 10.1016/j.fuel.2018.02.110.

- [61] L. O. Williams, "Fossil fuels," in *An End to Global Warming*, Elsevier, 2002, pp. 1–31.
- [62] M. J. K. Ahmed and M. Ahmaruzzaman, "Adsorptive desulfurization of feed diesel using chemically impregnated coconut coir waste," *Int. J. Environ. Sci. Technol.*, vol. 12, no. 9, pp. 2847–2856, 2015, doi: 10.1007/s13762-014-0654-4.
- [63] A. Baysal and S. Akman, "A practical method for the determination of sulphur in coal samples by high-resolution continuum source flame atomic absorption spectrometry," *Talanta*, vol. 85, no. 5, pp. 2662–2665, 2011, doi: 10.1016/j.talanta.2011.08.038.
- [64] K. Sun Yoo and L. M. Pike, "Determination of flavor precursor compound salk(en)yl-L-cysteine sulfoxides by an HPLC method and their distribution in allium species," *Sci. Hortic. (Amsterdam).*, vol. 75, no. 1–2, pp. 1–10, 1998, doi: 10.1016/S0304-4238(98)00107-1.
- [65] R. Kubec, M. Svobodová, and J. Velíšek, "Gas chromatographic determination of S-alk(en)ylcysteine sulfoxides," *J. Chromatogr. A*, vol. 862, no. 1, pp. 85–94, 1999, doi: 10.1016/S0021-9673(99)00902-4.
- [66] J. E. Lancaster and K. E. Kelly, "Quantitative analysis of the S-alk(en)yl-L-cysteine sulphoxides in onion (Allium cepa L.)," *J. Sci. Food Agric.*, vol. 34, no. 11, pp. 1229–1235, 1983, doi: 10.1002/jsfa.2740341111.
- [67] A. A. Bele and A. Khale., "an overview on thin layer chromatography Archana A. Bele* and Anubha Khale H. K. College of Pharmacy, Jogeshwari (W), Mumbai, Maharashtra, India," *J. Pharm. Sci.*, vol. 2, no. 2, pp. 256–267, 2011, doi: http://dx.doi.org/10.13040/IJPSR.0975-8232.2(2).256-67.
- [68] D. Harvey, Chromatographic and Electrophoretic Methods," *Anal. Chem. 2.1*, pp. 747–846, 2009.
- [69] S. Fajardo, García-Galvan, F. R., V. Barranco, J. C. Galvan, and S. F. Batlle, "We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists TOP 1 %," *Intech*, vol. i, no. tourism, p. 13, 2016, doi: http://dx.doi.org/10.5772/57353.

- [70] C. R. Wilson, "Methods for Analysis of Gastrointestinal Toxicants," *Compr. Toxicol. Second Ed.*, vol. 10, pp. 145–152, 2010, doi: 10.1016/B978-0-08-046884-6.00845-9.
- [71] H. Al-Rabiah, L. Al-Ostad, T. Al-Shamary, A. Al-Hendi, H. Al-Henayyan, and R. Khazemi, "Sulfur speciation of Kuwaiti crude oils fractions using twodimensional gas chromatography," *Arab. J. Chem.*, vol. 13, no. 2, pp. 3615– 3623, 2020, doi: 10.1016/j.arabjc.2019.11.007.
- [72] B. J. Pollo, G. L. Alexandrino, F. Augusto, and L. W. Hantao, "The impact of comprehensive two-dimensional gas chromatography on oil & gas analysis: Recent advances and applications in petroleum industry," *TrAC Trends Anal. Chem.*, vol. 105, pp. 202–217, 2018, doi: 10.1016/j.trac.2018.05.007.
- [73] W. E. Rudzinski and V. Rai, "Detection of polyaromatic sulfur heterocycles in crude oil using postcolumn addition of tropylium and tandem mass spectrometry," *Energy and Fuels*, vol. 19, no. 4, pp. 1611–1618, 2005, doi: 10.1021/ef0400666.
- [74] E. Tjabadi and N. Mketo, "Recent developments for spectrometric, chromatographic and electroanalytical determination of the total sulphur and halogens in various matrices," *TrAC - Trends Anal. Chem.*, vol. 118, pp. 207– 222, 2019, doi: 10.1016/j.trac.2019.05.033.
- [75] W. Wardencki, "Problems with the determination of environmental sulphur compounds by gas chromatography," *J. Chromatogr. A*, vol. 793, no. 1, pp. 1– 19, 1998, doi: 10.1016/S0021-9673(97)00997-7.
- [76] M. Yoo, S. Kim, S. Lee, and D. Shin, "Validated HPLC method and temperature stabilities for oil-soluble organosulfur compounds in garlic macerated oil," *J. Chromatogr. Sci.*, vol. 52, no. 10, pp. 1165–1172, 2014, doi: 10.1093/chromsci/bmt166.
- [77] C. Duyck, N. Miekeley, T. C. O. Fonseca, P. Szatmari, and E. V. D. S. Neto, "Trace element distributions in biodegraded crude oils and fractions from the Potiguar Basin, Brazil," *J. Braz. Chem. Soc.*, vol. 19, no. 5, pp. 978–986, 2008, doi: 10.1590/S0103-50532008000500025.

- [78] M. G. R. Vale *et al.*, "Method development for the determination of nickel in petroleum using line-source and high-resolution continuum-source graphite furnace atomic absorption spectrometry," *Microchem. J.*, vol. 77, no. 2, pp. 131–140, 2004, doi: 10.1016/j.microc.2004.02.007.
- [79] G. Parfene and G. Bahrim, "Evaluation of bioactive compounds in extracts obtained from three romanian marine algae species," *Rom Biotechnol Lett*, vol. 16, no. 6, pp. 71–78, 2011.
- [80] Q. Wang, T. Zhang, S. Zhang, Y. Fan, and B. Chen, "Extractive desulfurization of fuels using trialkylamine-based protic ionic liquids," *Sep. Purif. Technol.*, vol. 231, pp. 614–622, 2020, doi: 10.1016/j.seppur.2019.115923.
- [81] M. das G. A. Korn *et al.*, "Atomic spectrometric methods for the determination of metals and metalloids in automotive fuels - A review," *Talanta*, vol. 73, no. 1, pp. 1–11, 2007, doi: 10.1016/j.talanta.2007.03.036.
- [82] W. Yang, J. F. Casey, and Y. Gao, "A new sample preparation method for crude or fuel oils by mineralization utilizing single reaction chamber microwave for broader multi-element analysis by ICP techniques," *Fuel*, vol. 206, pp. 64– 79, 2017, doi: 10.1016/j.fuel.2017.05.084.
- [83] W. B. Wilson, H. V. Hayes, A. D. Campiglia, and S. A. Wise, "Qualitative characterization of three combustion-related standard reference materials for polycyclic aromatic sulfur heterocycles and their alkyl-substituted derivatives via normal-phase liquid chromatography and gas chromatography/mass spectrometry," *Anal. Bioanal. Chem.*, vol. 410, no. 17, pp. 4177–4188, 2018, doi: 10.1007/s00216-018-1065-z.
- [84] C. López García, M. Becchi, M. F. Grenier-Loustalot, O. Païsse, and R. Szymanski, "Analysis of aromatic sulfur compounds in gas oils using GC with sulfur chemiluminescence detection and high-resolution MS," *Anal. Chem.*, vol. 74, no. 15, pp. 3849–3857, 2002, doi: 10.1021/ac011190e.
- [85] G. A. Depauw and G. F. Froment, "Molecular analysis of the sulphur components in a light cycle oil of a catalytic cracking unit by gas chromatography with mass spectrometric and atomic emission detection," *J. Chromatogr. A*, vol. 761, no. 1–2, pp. 231–247, 1997, doi: 10.1016/S0021-

9673(96)00819-9.

- [86] R. Hua, "Analysis of sulfur-containing compounds in crude oils by comprehensive two-dimensional gas chromatography with sulfur chemiluminescence detection," *J.Sep. Sci*, pp. 691–698, 2004, doi: 10.1002/jssc.200301631.
- [87] B. M. F. Ávila, V. B. Pereira, A. O. Gomes, and D. A. Azevedo, "Speciation of organic sulfur compounds using comprehensive two-dimensional gas chromatography coupled to time-of-flight mass spectrometry : A powerful tool for petroleum refining," vol. 126, pp. 188–193, 2014, doi: 10.1016/j.fuel.2014.02.055.
- [88] WHO Regional Office for Europe, "Chapter 5.7 Dichloromethane," Air Qual. Guidel., vol. 2, no. Dcm, pp. 1–2, 2000, [Online]. Available: http://www.euro.who.int/__data/assets/pdf_file/0013/123061/AQG2ndEd_5_7D ichloromethane.pdf?ua=1.
- [89] W. B. Wilson, H. V. Hayes, L. C. Sander, A. D. Campiglia, and S. A. Wise, "Qualitative characterization of SRM 1597a coal tar for polycyclic aromatic hydrocarbons and methyl-substituted derivatives via normal-phase liquid chromatography and gas chromatography/mass spectrometry," *Anal. Bioanal. Chem.*, vol. 409, no. 21, pp. 5171–5183, 2017, doi: 10.1007/s00216-017-0464x.
- [90] R. Hua *et al.*, "Determination of sulfur-containing compounds in diesel oils by comprehensive two-dimensional gas chromatography with a sulfur chemiluminescence detector," *J. Chromatogr. A*, vol. 1019, no. 1–2, pp. 101– 109, 2003, doi: 10.1016/j.chroma.2003.08.048.
- [91] I. P. Afanas'ev, T. A. Gorshkova, S. A. Arystanbekova, M. S. Lapina, and A. B. Volynskii, "Determination of individual sulfur-containing compounds in gas condensate and petrol by gas chromatography," *J. Anal. Chem.*, vol. 72, no. 10, pp. 1095–1101, 2017, doi: 10.1134/S1061934817100021.
- [92] G. Zhu, M. Wang, and T. Zhang, "Identification of polycyclic sulfides hexahydrodibenzothiophenes and their implications for heavy oil accumulation in ultra-deep strata in Tarim Basin," *Mar. Pet. Geol.*, vol. 78, pp. 439–447,

2016, doi: 10.1016/j.marpetgeo.2016.09.027.

- [93] L. Mahé, T. Dutriez, M. Courtiade, D. Thiébaut, H. Dulot, and F. Bertoncini, "Global approach for the selection of high temperature comprehensive twodimensional gas chromatography experimental conditions and quantitative analysis in regards to sulfur-containing compounds in heavy petroleum cuts," *J. Chromatogr. A*, vol. 1218, no. 3, pp. 534–544, 2011, doi: 10.1016/j.chroma.2010.11.065.
- [94] U. Nylén, J. F. Delgado, S. Järås, and M. Boutonnet, "Characterization of alkylated aromatic sulphur compounds in light cycle oil from hydrotreated vacuum gas oil using GC-SCD," *Fuel Process. Technol.*, vol. 86, no. 2, pp. 223–234, 2004, doi: 10.1016/j.fuproc.2004.03.001.
- [95] J. Heilmann and K. G. Heumann, "Development of a species-specific isotope dilution GC-ICP-MS method for the determination of thiophene derivates in petroleum products," *Anal. Bioanal. Chem.*, vol. 390, no. 2, pp. 643–653, 2008, doi: 10.1007/s00216-007-1574-7.
- [96] L. L. P. Van Stee, J. Beens, R. J. J. Vreuls, and U. A. T. Brinkman, "Comprehensive two-dimensional gas chromatography with atomic emission detection and correlation with mass spectrometric detection: Principles and application in petrochemical analysis," *J. Chromatogr. A*, vol. 1019, no. 1–2, pp. 89–99, 2003, doi: 10.1016/S0021-9673(03)01301-3.
- [97] B. Mitrevski, M. W. Amer, A. L. Chaffee, and P. J. Marriott, "Evaluation of comprehensive two-dimensional gas chromatography with flame photometric detection: Potential application for sulfur speciation in shale oil," *Anal. Chim. Acta*, vol. 803, pp. 174–180, 2013, doi: 10.1016/j.aca.2013.07.021.
- [98] B. Zappacosta *et al.*, "HPLC analysis of some sulphur compounds in saliva: Comparison between healthy subjects and periodontopathic patients," *Clin. Chim. Acta*, vol. 338, no. 1–2, pp. 57–60, 2003, doi: 10.1016/j.cccn.2003.07.019.
- [99] D. W. Jacobsen *et al.*, "Rapid HPLC determination of total homocysteine and other thiols in serum and plasma: Sex differences and correlation with cobalamin and folate concentrations in healthy subjects," *Clin. Chem.*, vol. 40,

no. 6, pp. 873–881, 1994, doi: 10.1093/clinchem/40.6.873.

- [100] M. Petrović, S. Gonzalez, and D. Barceló, "Analysis and removal of emerging contaminants in wastewater and drinking water," *TrAC - Trends Anal. Chem.*, vol. 22, no. 10, pp. 685–696, 2003, doi: 10.1016/S0165-9936(03)01105-1.
- [101] C. I. C. Silvestre, J. L. M. Santos, J. L. F. C. Lima, and E. A. G. Zagatto,
 "Liquid-liquid extraction in flow analysis: A critical review," *Anal. Chim. Acta*, vol. 652, no. 1–2, pp. 54–65, 2009, doi: 10.1016/j.aca.2009.05.042.
- [102] V. P. Jofré, M. V. Assof, M. L. Fanzone, H. C. Goicoechea, L. D. Martínez, and M. F. Silva, "Optimization of ultrasound assisted-emulsification-dispersive liquid-liquid microextraction by experimental design methodologies for the determination of sulfur compounds in wines by gas chromatography-mass spectrometry," *Anal. Chim. Acta*, vol. 683, no. 1, pp. 126–135, 2010, doi: 10.1016/j.aca.2010.10.010.
- [103] I. Al-zahrani, C. Basheer, and T. Htun, "Application of liquid-phase microextraction for the determination of sulfur compounds in crude oil and diesel," *J. Chromatogr. A*, vol. 1330, pp. 97–102, 2014, doi: 10.1016/j.chroma.2014.01.015.
- [104] H. M. Al-Saidi and A. A. A. Emara, "The recent developments in dispersive liquid-liquid microextraction for preconcentration and determination of inorganic analytes," *J. Saudi Chem. Soc.*, vol. 18, no. 6, pp. 745–761, 2014, doi: 10.1016/j.jscs.2011.11.005.
- [105] J. M. Kokosa, "Advances in solvent-microextraction techniques," *TrAC Trends Anal. Chem.*, vol. 43, pp. 2–13, Feb. 2013, doi: 10.1016/J.TRAC.2012.09.020.
- [106] A. Sarafraz-Yazdi and A. Amiri, "Liquid-phase microextraction," *TrAC Trends Anal. Chem.*, vol. 29, no. 1, pp. 1–14, 2010, doi: 10.1016/j.trac.2009.10.003.
- [107] J. Xiong and B. Hu, "Comparison of hollow fiber liquid phase microextraction and dispersive liquid-liquid microextraction for the determination of organosulfur pesticides in environmental and beverage samples by gas chromatography with flame photometric detection," *J. Chromatogr. A*, vol. 1193, no. 1–2, pp. 7–18, 2008, doi: 10.1016/j.chroma.2008.03.072.

- [108] A. Mohamed, A. Alsharif, G. Tan, Y. Choo, and A. Lawal, "Ef fi ciency of Hollow Fiber Liquid-Phase Microextraction Chromatography Methods in the Separation of Organic Compounds : A Review," vol. 55, no. 3, pp. 378–391, 2017, doi: 10.1093/chromsci/bmw188.
- [109] S. Shariati, Y. Yamini, and M. Rezaee, "Dispersive liquid-liquid microextraction for the preconcentration and determination of some organic sulfur compounds in aqueous samples," *Monatshefte fur Chemie*, vol. 142, no. 6, pp. 555–560, 2011, doi: 10.1007/s00706-011-0483-9.
- [110] B. Mokhtari, N. Dalali, and K. Pourabdollah, "Preconcentration and determination of short chain mercaptans in the produced water by dispersive liquid-liquid microextraction and gas chromatography," *J. Chil. Chem. Soc.*, vol. 57, no. 4, pp. 1428–1431, 2012, doi: 10.4067/S0717-97072012000400019.
- [111] Q. Xiao, C. Yu, J. Xing, and B. Hu, "Comparison of headspace and direct single-drop microextraction and headspace solid-phase microextraction for the measurement of volatile sulfur compounds in beer and beverage by gas chromatography with flame photometric detection," *J. Chromatogr. A*, vol. 1125, no. 1, pp. 133–137, 2006, doi: 10.1016/j.chroma.2006.06.096.
- [112] N. Moreira, P. Guedes De Pinho, and I. Vasconcelos, "Method for analysis of heavy sulphur compounds using gas chromatography with flame photometric detection," *Anal. Chim. Acta*, vol. 513, no. 1, pp. 183–189, 2004, doi: 10.1016/j.aca.2003.12.041.
- [113] Z. Qriouet, Z. Qmichou, N. Bouchoutrouch, H. Mahi, Y. Cherrah, and H. Sefrioui, "Analytical Methods Used for the Detection and Quantification of Benzodiazepines," *J. Anal. Methods Chem.*, vol. 2019, 2019, doi: 10.1155/2019/2035492.
- [114] N. Fontanals, R. M. Marcé, and F. Borrull, "Materials for solid-phase extraction of organic compounds," *Separations*, vol. 6, no. 4, 2019, doi: 10.3390/separations6040056.
- [115] K. M. Dimpe and P. N. Nomngongo, "Current sample preparation methodologies for analysis of emerging pollutants in different environmental

matrices," *TrAC - Trends Anal. Chem.*, vol. 82, pp. 199–207, 2016, doi: 10.1016/j.trac.2016.05.023.

- [116] S. G. Mössner and S. A. Wise, "Determination of polycyclic aromatic sulfur heterocycles in fossil fuel-related samples," *Anal. Chem.*, vol. 71, no. 1, pp. 58–69, 1999, doi: 10.1021/ac980664f.
- [117] Y. Su, S. Xia, R. Wang, and L. Xiao, Phytohormonal quantification based on biological principles, *In hormone metabolism and signaling in plants*, 2017.
- [118] Q. Gao, D. Luo, J. Ding, and Y. Q. Feng, "Rapid magnetic solid-phase extraction based on magnetite/silica/poly(methacrylic acid-co-ethylene glycol dimethacrylate) composite microspheres for the determination of sulfonamide in milk samples," *J. Chromatogr. A*, vol. 1217, no. 35, pp. 5602–5609, 2010, doi: 10.1016/j.chroma.2010.06.067.
- [119] G. Domingos, C. Katielli, R. Bueno, L. Machado, and D. Carvalho, "A cleanup method using solid phase extraction for the determination of organosulfur compounds in petroleum asphalt cements," vol. 202, pp. 206–215, 2017, doi: 10.1016/j.fuel.2017.04.020.
- [120] J. T. Andersson, "Separation methods in the analysis of polycyclic aromatic sulfur heterocycles in handbook of analytical separation, wolfgang Kleibohmer, Elsivier science B.V" vol. 3, pp. 96–98, 2001.
- [121] J. Pawliszyn, "Theory of Solid-Phase Microextraction," vol. 38, no. July, pp. 270–278, 2000.
- [122] M. A. Mottaleb, M. J. Meziani, and M. R. Islam, "Solid-Phase Microextraction and its Application to Natural Products," *Encycl. Anal. Chem.*, no. January 2014, pp. 1–23, 2014, doi: 10.1002/9780470027318.a9905.
- [123] F. Pelusio *et al.*, "Headspace Solid-Phase Microextraction Analysis of Volatile Organic Sulfur Compounds in Black and White Truffle Aroma," *J. Agric. Food Chem.*, vol. 43, no. 8, pp. 2138–2143, 1995, doi: 10.1021/jf00056a034.
- [124] F. Lestremau, F. A. T. Andersson, V. Desauziers, and J. L. Fanlo, "Evaluation of solid-phase microextraction for time-weighted average sampling of volatile sulfur compounds at ppb concentrations," *Anal. Chem.*, vol. 75, no. 11, pp.
2626–2632, 2003, doi: 10.1021/ac034124g.

- [125] C. Haberhauer-Troyer, E. Rosenberg, and M. Grasserbauer, "Evaluation of solid-phase microextraction for sampling of volatile organic sulfur compounds in air for subsequent gas chromatographic analysis with atomic emission detection," *J. Chromatogr. A*, vol. 848, no. 1–2, pp. 305–315, 1999, doi: 10.1016/S0021-9673(99)00459-8.
- [126] P. G. Hill and R. M. Smith, "Determination of sulphur compounds in beer using headspace solid-phase microextraction and gas chromatographic analysis with pulsed flame photometric detection," *J. Chromatogr. A*, vol. 872, no. 1–2, pp. 203–213, 2000, doi: 10.1016/S0021-9673(99)01307-2.
- [127] P. Fuchsmann, M. T. Stern, Y. A. Brügger, and K. Breme, "Olfactometry Profiles and Quantitation of Volatile Sulfur Compounds of Swiss Tilsit Cheeses," *J. Agric. Food Chem.*, vol. 63, no. 34, pp. 7511–7521, 2015, doi: 10.1021/acs.jafc.5b02536.
- [128] N. Luna, L. Zelong, and T. Songbai, "Identification and characterization of sulfur compounds in straight-run diesel using comprehensive two-dimensional GC coupled with TOF MS," *China Pet. Process. Petrochemical Technol.*, vol. 16, no. 3, pp. 10–18, 2014.
- [129] X. Lu, C. Fan, J. Shang, J. Deng, and H. Yin, "Headspace solid-phase microextraction for the determination of volatile sulfur compounds in odorous hyper-eutrophic freshwater lakes using gas chromatography with flame photometric detection," *Microchem. J.*, vol. 104, pp. 26–32, 2012, doi: 10.1016/j.microc.2012.04.001.
- [130] D. A. Locatelli, J. C. Altamirano, J. M. Luco, R. Norlin, and A. B. Camargo,
 "Solid phase microextraction coupled to liquid chromatography. Analysis of organosulphur compounds avoiding artifacts formation," *Food Chem.*, vol. 157, pp. 199–204, 2014, doi: 10.1016/j.foodchem.2014.02.010.
- [131] O. Calvo-Gómez, J. Morales-López, and M. G. López, "Solid-phase microextraction-gas chromatographic-mass spectrometric analysis of garlic oil obtained by hydrodistillation," *J. Chromatogr. A*, vol. 1036, no. 1, pp. 91–93, 2004, doi: 10.1016/j.chroma.2004.02.072.

- [132] A. T. Nielsen and S. Jonsson, "Quantification of volatile sulfur compounds in complex gaseous matrices by solid-phase microextraction," *J. Chromatogr. A*, vol. 963, no. 1–2, pp. 57–64, 2002, doi: 10.1016/S0021-9673(02)00556-3.
- [133] R. A. Murray, "Limitations to the use of solid-phase microextraction for quantitation of mixtures of volatile organic sulfur compounds," *Anal. Chem.*, vol. 73, no. 7, pp. 1646–1649, 2001, doi: 10.1021/ac001176m.
- [134] W. B. Studabaker, K. J. Puckett, K. E. Percy, and M. S. Landis, "Determination of polycyclic aromatic hydrocarbons, dibenzothiophene, and alkylated homologs in the lichen Hypogymnia physodes by gas chromatography using single quadrupole mass spectrometry and time-of-flight mass spectrometry," *J. Chromatogr. A*, vol. 1492, pp. 106–116, 2017, doi: 10.1016/j.chroma.2017.02.051.
- [135] P. A. Vazquez-Landaverde, J. A. Torres, and M. C. Qian, "Quantification of trace volatile sulfur compounds in milk by solid-phase microextraction and gas chromatography-pulsed flame photometric detection," *J. Dairy Sci.*, vol. 89, no. 8, pp. 2919–2927, 2006, doi: 10.3168/jds.S0022-0302(06)72564-4.
- [136] S. Rochat, J. Y. de Saint Laumer, and A. Chaintreau, "Analysis of sulfur compounds from the in-oven roast beef aroma by comprehensive twodimensional gas chromatography," *J. Chromatogr. A*, vol. 1147, no. 1, pp. 85– 94, 2007, doi: 10.1016/j.chroma.2007.02.039.
- [137] E. Kabir and K. H. Kim, "Use of solid phase microextraction (SPME) in the analysis of the reduced sulfur compounds (RSC) and its experimental limitations," *Microchem. J.*, vol. 103, pp. 42–48, 2012, doi: 10.1016/j.microc.2012.01.005.
- [138] C. Salgado-Petinal, R. Alzaga, C. García-Jares, M. Llompart, and J. M. Bayona, "Low part per trillion determination of reactive alkanethiols in wastewater by in situ derivatization-solid-phase microextraction followed by GC/MS," *Anal. Chem.*, vol. 77, no. 18, pp. 6012–6018, 2005, doi: 10.1021/ac050685r.
- [139] R. López, A. C. Lapeña, J. Cacho, and V. Ferreira, "Quantitative determination of wine highly volatile sulfur compounds by using automated headspace solid-

phase microextraction and gas chromatography-pulsed flame photometric detection. Critical study and optimization of a new procedure," *J. Chromatogr. A*, vol. 1143, no. 1–2, pp. 8–15, 2007, doi: 10.1016/j.chroma.2006.12.053.

- [140] Y. Fang and M. C. Qian, "Sensitive quantification of sulfur compounds in wine by headspace solid-phase microextraction technique," *J. Chromatogr. A*, vol. 1080, no. 2, pp. 177–185, 2005, doi: 10.1016/j.chroma.2005.05.024.
- [141] L. Mateo-Vivaracho, V. Ferreira, and J. Cacho, "Automated analysis of 2methyl-3-furanthiol and 3-mercaptohexyl acetate at ng L-1 level by headspace solid-phase microextracion with on-fibre derivatisation and gas chromatography-negative chemical ionization mass spectrometric determination," *J. Chromatogr. A*, vol. 1121, no. 1, pp. 1–9, 2006, doi: 10.1016/j.chroma.2006.04.029.
- [142] P. M. Kalenga, "Determination and Characterization of Sulphur in South African Coal," *Unpubl. MSc Thesis*, no. 367937, p. 142, 2011.
- [143] G. Gryglewicz, P. Rutkowski, and J. Yperman, "Characterization of sulfur compounds in supercritical coal extracts by gas chromatography-mass spectrometry," *Fuel Process. Technol.*, vol. 77–78, pp. 167–172, 2002, doi: 10.1016/S0378-3820(02)00046-2.
- [144] M. E. MacHado, F. Cappelli Fontanive, J. V. De Oliveira, E. B. Caramão, and C. Alcaraz Zini, "Identification of organic sulfur compounds in coal bitumen obtained by different extraction techniques using comprehensive twodimensional gas chromatography coupled to time-of-flight mass spectrometric detection," *Anal. Bioanal. Chem.*, vol. 401, no. 8, pp. 2433–2444, 2011, doi: 10.1007/s00216-011-5171-4.
- [145] J. T. Andersson, T. Schade, and H. Müller, "Separation of aromatic sulfur heterocycles from aromatic hydrocarbons by use of a palladium ion complex," ACS Div. Fuel Chem. Prepr., vol. 47, no. 2, pp. 697–698, 2002.
- [146] K. Sripada and J. T. Andersson, "Liquid chromatographic properties of aromatic sulfur heterocycles on a Pd(II)-containing stationary phase for petroleum analysis," *Anal. Bioanal. Chem.*, vol. 382, no. 3, pp. 735–741, 2005, doi: 10.1007/s00216-004-3026-y.

- [147] N. Ghaloum, G. Michael, and Z. Khan, "Separation of Sulphur Compounds From Vacuum Gas Oil Distillates By Ligand Exchange Chromatography," *J. Liq. Chromatogr. Relat. Technol.*, vol. 25, no. 9, pp. 1409–1420, 2002, doi: 10.1081/jlc-120004756.
- [148] G. Vasapollo *et al.*, "Molecularly imprinted polymers: Present and future prospective," *Int. J. Mol. Sci.*, vol. 12, no. 9, pp. 5908–5945, 2011, doi: 10.3390/ijms12095908.
- [149] M. R. Ras, F. Borrull, and R. M. Marcé, "Determination of volatile organic sulfur compounds in the air at sewage management areas by thermal desorption and gas chromatography-mass spectrometry," *Talanta*, vol. 74, no. 4, pp. 562–569, 2008, doi: 10.1016/j.talanta.2007.06.017.
- [150] E. Frerot, A. Bagnoud, and E. Cicchetti, "Quantification of hydrogen sulfide and methanethiol and the study of their scavenging by biocides of the isothiazolone family," *Chempluschem*, vol. 79, no. 1, pp. 77–82, 2014, doi: 10.1002/cplu.201300234.
- [151] G. Gryglewicz and P. Rutkowski, "Identification of organic sulfur compounds in supercritical extracts from. Polish lignite," *Energy and Fuels*, vol. 15, no. 1, pp. 8–13, 2001, doi: 10.1021/ef000085p.
- [152] S. Bose, B. Laha, and S. Banerjee, "Quantification of allicin by high performance liquid chromatography-ultraviolet analysis with effect of postultrasonic sound and microwave radiation on fresh garlic cloves," *Pharmacogn. Mag.*, vol. 10, no. 38, p. 288, 2014, doi: 10.4103/0973-1296.133279.
- [153] J. C. Zhu, Y. W. Niu, and Z. B. Xiao, "Characterization of important sulfur and nitrogen compounds in Lang baijiu by application of gas chromatographyolfactometry, flame photometric detection, nitrogen phosphorus detector and odor activity value," *Food Res. Int.*, vol. 131, no. January, p. 109001, 2020, doi: 10.1016/j.foodres.2020.109001.
- [154] N. S. Kanaki and M. Rajani, "Development and validation of a thin-layer chromatography-densitometric method for the quantitation of alliin from garlic (allium sativum) and its formulations," *J. AOAC Int.*, vol. 88, no. 5, pp. 1568– 1570, 2005, doi: 10.1093/jaoac/88.5.1568.

- [155] N. A. Siddiqui, R. A. Mothana, and P. Alam, "Quantitative determination of alliin in dried garlic cloves and products by high-performance thin-layer chromatography," *Trop. J. Pharm. Res.*, vol. 15, no. 8, pp. 1759–1765, 2016, doi: 10.4314/tjpr.v15i8.23.
- [156] X. Song *et al.*, "Characterization of key aroma-active sulfur-containing compounds in Chinese Laobaigan Baijiu by gas chromatography-olfactometry and comprehensive two-dimensional gas chromatography coupled with sulfur chemiluminescence detection," *Food Chem.*, vol. 297, no. May, p. 124959, 2019, doi: 10.1016/j.foodchem.2019.124959.
- [157] L. Gîtin, R. Dinicə, C. Neagu, and L. Dumitrascu, "Sulfur compounds identification and quantification from Allium spp. fresh leaves," *J. Food Drug Anal.*, vol. 22, no. 4, pp. 425–430, 2014, doi: 10.1016/j.jfda.2014.04.002.
- [158] H. Y. Hijazi and C. S. Bottaro, "Molecularly imprinted polymer thin-film as a micro-extraction adsorbent for selective determination of trace concentrations of polycyclic aromatic sulfur heterocycles in seawater," *J. Chromatogr. A*, no. xxxx, 2020, doi: 10.1016/j.chroma.2019.460824.
- [159] M. Mestres, O. Busto, and J. Guasch, "Chromatographic analysis of volatile sulphur compounds in wines using the static headspace technique with flame photometric detection," *J. Chromatogr. A*, vol. 773, no. 1–2, pp. 261–269, 1997, doi: 10.1016/S0021-9673(97)00188-X.
- [160] V. Carrascon, I. Ontañón, M. Bueno, and V. Ferreira, "Gas chromatographymass spectrometry strategies for the accurate and sensitive speciation of sulfur dioxide in wine," *J. Chromatogr. A*, vol. 1504, pp. 27–34, 2017, doi: 10.1016/j.chroma.2017.05.012.
- [161] E. Zambrzycka and B. Godlewska-Zyłkiewicz, "Determination of sulfur in food by high resolution continuum source flame molecular absorption spectrometry," *Spectrochim. Acta - Part B At. Spectrosc.*, vol. 101, pp. 234– 239, 2014, doi: 10.1016/j.sab.2014.08.041.
- [162] P. Q. Tranchida, M. Zoccali, I. Bonaccorsi, P. Dugo, L. Mondello, and G. Dugo,"The off-line combination of high performance liquid chromatography and comprehensive two-dimensional gas chromatography-mass spectrometry: A

powerful approach for highly detailed essential oil analysis," *J. Chromatogr. A*, vol. 1305, pp. 276–284, 2013, doi: 10.1016/j.chroma.2013.07.042.

- [163] D. Mao, H. Van De Weghe, L. Diels, N. De Brucker, R. Lookman, and G. Vanermen, "High-performance liquid chromatography fractionation using a silver-modified column followed by two-dimensional comprehensive gas chromatography for detailed group-type characterization of oils and oil pollutions," *J. Chromatogr. A*, vol. 1179, no. 1, pp. 33–40, 2008, doi: 10.1016/j.chroma.2007.09.085.
- [164] A. Ali *et al.*, "Synthesis, characterization, applications, and challenges of iron oxide nanoparticles," *Nanotechnol. Sci. Appl.*, vol. 9, pp. 49–67, 2016, doi: 10.2147/NSA.S99986.
- [165] P. Tartaj, S. Veintemillas-Verdaguer, T. Gonzalez-Carreño, and C. J. Serna, "Preparation of magnetic nanoparticles for applications in biomedicine," *Magn. Nanoparticles Biosensing Med.*, pp. 52–67, 2019, doi: 10.1017/9781139381222.003.
- [166] S. D. lancu *et al.*, "Assessment of gold-coated iron oxide nanoparticles as negative T2 contrast agent in small animal MRI studies," *Int. J. Nanomedicine*, vol. 15, pp. 4811–4824, 2020, doi: 10.2147/IJN.S253184.
- [167] J. P. Jolivet, P. Belleville, E. Tronc, and J. Livage, "Influence of Fe(II) on the formation of the spinel iron oxide in alkaline medium," *Clays Clay Miner.*, vol. 40, no. 5, pp. 531–539, 1992, doi: 10.1346/CCMN.1992.0400506.
- [168] Q. H. Lu, K. L. Yao, D. Xi, Z. L. Liu, X. P. Luo, and Q. Ning, "Synthesis and characterization of composite nanoparticles comprised of gold shell and magnetic core / cores," *J.Magn.Magn*, vol. 301, pp. 44–49, 2006, doi: 10.1016/j.jmmm.2005.06.007.
- [169] N. S. Elbialy, M. M. Fathy, and W. M. Khalil, "Doxorubicin loaded magnetic gold nanoparticles for in vivo targeted drug delivery," *Int. J. Pharm.*, vol. 490, no. 1–2, pp. 190–199, 2015, doi: 10.1016/j.ijpharm.2015.05.032.
- [170] C. K. Lo, D. Xiao, and M. M. F. Choi, "Homocysteine-protected gold-coated magnetic nanoparticles: Synthesis and characterisation," *J. Mater. Chem.*, vol.

17, no. 23, pp. 2418–2427, 2007, doi: 10.1039/b617500g.

- [171] X. An, W. Chai, X. Deng, H. Chen, and G. Ding, "A bioinspired polydopamine approach toward the preparation of gold-modified magnetic nanoparticles for the magnetic solid-phase extraction of steroids in multiple samples," *J. Sep. Sci.*, vol. 41, no. 13, pp. 2774–2782, 2018, doi: 10.1002/jssc.201800080.
- [172] M. Almeida, S. Luis, C. Ferreira, C. Galvão, A. Maria, and G. Souza, "Talanta Simultaneous optimization of multiple responses and its application in Analytical Chemistry – A review," *Talanta*, vol. 194, no. October 2018, pp. 941–959, 2019, doi: 10.1016/j.talanta.2018.10.088.
- [173] P. N. Nomngongo and J. C. Ngila, "Multivariate optimization of dual-bed solid phase extraction for preconcentration of Ag , AI , As and Cr in gasoline prior to inductively coupled plasma optical emission spectrometric determination," *FUEL*, vol. 139, pp. 285–291, 2015, doi: 10.1016/j.fuel.2014.08.046.
- [174] M. Ndilimeke and K. Mogolodi, "Ultrasonic assisted magnetic solid phase extraction based on the use of magnetic waste-tyre derived activated carbon modified with methyltrioctylammonium chloride adsorbent for the preconcentration and analysis of non-steroidal anti- inflammatory drugs in," *Arab. J. Chem.*, vol. 14, no. 9, p. 103329, 2021, doi: 10.1016/j.arabjc.2021.103329.
- [175] B. Maddi, E. Panisko, K. Albrecht, and D. Howe, "Qualitative characterization of the aqueous fraction from hydrothermal liquefaction of algae using 2D gas chromatography with time-of-flight mass spectrometry," *J. Vis. Exp.*, vol. 2016, no. 109, pp. 1–11, 2016, doi: 10.3791/53634.
- [176] "UV-Vis Spectroscopy: Principle, Strengths and Limitations and Applications | Technology Networks." [Online]. Available: https://www.technologynetworks.com/analysis/articles/uv-vis-spectroscopyprinciple-strengths-and-limitations-and-applications-349865.
- [177] M. Tare, S. Pilani, O. R. Puli, and A. Singh, "Drosophila adult eye model to teach Scanning Electron Microscopy in an undergraduate cell biology laboratory," no. December 2016, 2009.

- [178] M. D. Mccluskey, "Local Vibrational Modes of Impurities in Semiconductors Local vibrational modes of impurities in semiconductors," *J. Appl.Phys* no. April 2000, 2013, doi: 10.1063/1.372453.
- [179] S. E. Bohndiek *et al.*, "A CMOS active pixel sensor system for laboratorybased x-ray diffraction studies of biological tissue," *Phys. Med. Biol.*, vol. 53, no. 3, pp. 655–672, 2008, doi: 10.1088/0031-9155/53/3/010.
- [180] L. C. Kelly, "Detection and identification of sulfur compounds in an Australian jet fuel [electronic resource] / Lance C. Kelly and Paul Rawson," *Tech. Note* (*Defence Sci. Technol. Organ. Online*); *DSTO-TN-0956*, 2010, [Online]. Available: http://nla.gov.au/nla.arc-24764.
- [181] A. Vetere, D. Pröfrock, and W. Schrader, "Quantitative and Qualitative Analysis of Three Classes of Sulfur Compounds in Crude Oil," *Angew. Chemie* - *Int. Ed.*, vol. 56, no. 36, pp. 10933–10937, 2017, doi: 10.1002/anie.201703205.
- [182] I. Aloisi, M. Zoccali, P. Q. Tranchida, and L. Mondello, "Analysis of organic sulphur compounds in coal tar by using comprehensive two-dimensional gas chromatography-high resolution time-of-flight mass spectrometry," *Separations*, vol. 7, no. 2, pp. 3–11, 2020, doi: 10.3390/separations7020026.
- [183] N. S. Elbialy, M. M. Fathy, and W. M. Khalil, "Preparation and characterization of magnetic gold nanoparticles to be used as doxorubicin nanocarriers," *Phys. Medica*, vol. 30, no. 7, pp. 843–848, 2014, doi: 10.1016/j.ejmp.2014.05.012.
- [184] T. T. Hien Pham, C. Cao, and S. J. Sim, "Application of citrate-stabilized goldcoated ferric oxide composite nanoparticles for biological separations," *J. Magn. Magn. Mater.*, vol. 320, no. 15, pp. 2049–2055, 2008, doi: 10.1016/j.jmmm.2008.03.015.
- [185] R. Stein *et al.*, "Synthesis and characterization of citrate-stabilized gold-coated superparamagnetic iron oxide nanoparticles for biomedical applications," *Molecules*, vol. 25, no. 19, 2020, doi: 10.3390/molecules25194425.
- [186] M. Ghorbani, H. Hamishehkar, N. Arsalani, and A. A. Entezami, "Preparation of thermo and pH-responsive polymer@Au/Fe3O4 core/shell nanoparticles as

a carrier for delivery of anticancer agent," *J. Nanoparticle Res.*, vol. 17, no. 7, pp. 1–13, 2015, doi: 10.1007/s11051-015-3097-z.

- [187] T. A. Ivandini, R. Sato, Y. Makide, A. Fujishima, and Y. Einaga,
 "Electrochemical Detection of Arsenic (III) Using Electrodes," *Anal. Chem.*, vol. 78, no. 18, pp. 6291–6298, 2006.
- [188] DoE, "Government Gazette Staatskoerant," Gov. Gaz., vol. 583, no. 37230, pp. 1–4, 2014, [Online]. Available: http://www.greengazette.co.za/pages/national-gazette-37230-of-17-january-2014-vol-583_20140117-GGN-37230-003.